

REVIEW ARTICLE

DRUG DELIVERY TO RETINA: A REVIEW

Shelke D. A.^a and Shirolkar S.^{a*}

(Received 06 July 2019) (Accepted 06 August 2019)

ABSTRACT

The drug delivery to posterior segment especially to retina of eye is difficult due to various barriers. The diseases affecting the retina of eye are increasing and hence there is need to develop approaches for drug delivery to retina. This review describes the anatomy of retina, barriers associated with it, and diseases of retina. The drug delivery to retina by systemic, topical, intravitreal injection, intravitreal implant along with advance nanotechnology based and transporter mediated drug delivery is discussed here. The recent technologies in retinal drug delivery are also discussed to give comprehensive recent information about retinal drug delivery.

Keywords: Retina, Retinal Drug delivery, Transporter, Nanotechnology, Intravitreal.

INTRODUCTION

Drug delivery to the posterior segment of eye is challenging. The retina is the primary target for different vascular, degenerative and proliferative disorders. Because of anatomical and physiological barriers associated with the retina, it remains a challenge for drug delivery to achieve continuous therapeutic concentrations of the drug at the retina. In case of chronic therapy, required in conditions like age related macular degeneration and diabetic retinopathy the challenge is reversed. The topical administration of drug in the lower cal-de-sac has been successful for treatment of diseases affecting the anterior segment of the eye. The cornea and conjunctiva are primary barriers after topical administration of ocular drugs. The other barriers like nasolacrimal drainage, protein binding, long diffusional path lengths, drug metabolism and lens barrier hinders the uptake of drug into the retina after topical administration. Systemic administration of drug to retina is also limited by blood ocular barriers namely blood aqueous and blood retinal barriers¹. The blood retinal barrier is formed by retinal pigmented epithelium (RPE) and endothelium of retinal vessels. The blood aqueous barrier includes endothelium of the iris vessels and non-pigmented layer of the ciliary epithelium². Due to the presence of these barriers, the achievement of therapeutic effect in the retina requires a higher systemic

dose but that can produce harmful side effects. Various local modes of administration like intravitreal, intracameral and periocular routes were also investigated to improve retinal drug delivery. The injection of drug into the anterior chamber is recognized as intracameral injections. By this route only up to 100 μ L volume can be administered³⁻⁶. These injections are used in management of anterior segment diseases and cataract surgery. However, significant concentrations of drug to the posterior segment cannot be delivered by this route.

Intravitreal injection is found to have wide clinical applications in treating vitreoretinal disorders. The first report on administration of this mode was in 1944. This route provides a higher concentration of drug to the neural retina since different barriers are not involved. But higher dose intravitreal injections resulted in higher local concentration of drug, which causes retinal toxicity e.g. gentamicin⁷. The repeated administration of injections through the intravitreal route results in retinal detachment. With increase in molecular weight, in vitreous the elimination half-lives of drugs also increase where as in plasma, protein and peptide drugs show shorter half-lives. Due to unique vitreal clearance property, macromolecules like vascular endothelial growth factor (VEGF) antibody and VEGF aptamer are under development for intravitreal administration⁸. Such drugs require repeated intravitreal administration hence they might increase the risk in treating chronic disorders.

^a Department of Pharmaceutics, Dr. D.Y.Patil Institute of Pharmaceutical Sciences and Research, Pimpri, Pune - 411018, Maharashtra, India

* For Correspondence E-mail: satishshirolkar@yahoo.com ; dshelke89@gmail.com

In spite of many advances in the field of formulation of ocular drug delivery, targeting drug to the posterior segment of the eye remained challenging due to different barriers in delivery. The increasing aging population and age-related retinal disease⁹, is worrying because it is increasing the burden to the global health care system. There is a clear and urgent requirement to develop smarter formulation strategies to overcome the barriers associated with the posterior eye and cater to this increasing patient need.

The intravitreal injection is currently the most effective and practical route for the treatment of retinal diseases but it has risk because of the poor rate of patient acceptability. There are many formulation strategies aiming to improve intravitreal delivery and efficacy while reducing the frequency of treatment¹⁰. There is need to gain better understanding of anatomy and physiology of the targets to achieve this goal by researchers. This review gives a comprehensive idea about the anatomy of the eye especially retina of the eye, the diseases associated with retina and drug delivery strategies used in the treatment of retinal disease. This review will explore the advances in drug delivery systems like nanoparticles, transporters and transporter mediated drug delivery approaches for the retina.

ANATOMY OF EYE AND RETINA

Structure of eye

Fig. 1 depicts structure of eye in a schematic form. The eye is composed of two major segments as the anterior segment and posterior segment divided by lens. The Posterior segment is of three major layers as outer sclera, middle choroid and inner retina. The vitreous chamber is main body filled with gel like vitreous humor surrounded by three layers. The outer sclera provides structural stability while middle choroid is highly vascular layer which provide nourishment to retina and metabolite clearance from the retina. The retina is the innermost and important layer composed of the complex neurosensory layer of the retina. The vitreous humor is about 4 mL and composed of water (99%), collogen, non-collagenous proteins and glycosaminoglycans. The vitreous humor has two key roles

(i) integrity of structure and (ii) transportation of nutrients to the retina and metabolites from the retina¹¹.

Structure of Retina

Fig. 2 represents structure of retina. The retina has two main parts Neural retina and Retinal pigmented

epithelium (RPE) and the space separating these two parts known as the subretinal space¹². The neural retina is also known as neuro sensory retina and is composed of a seven layered structure. The light enters from the Ganglionic cell layer (GCL) and penetrates all layers to reach the Rods and Cones. This signal in the form of a transduced signal is conducted out of the retina via the optic nerve to the brain and an image is formed.

The RPE is a single cell layer separating the outer surface of neural retina from choroid. The RPE has two portions apical portion and basolateral portion. The apical portion faces the neural retina and basolateral portion is next to the choroid. The apical portion is made up of numerous microvilli while the basolateral portion is made up of small convoluted foldings which increases the area of absorption for the choroid side. Together the RPE is the specific nutrient transport system which maintains the environment of retina.

The neural element portion of retina is separated from the blood supply by the endothelial cell of retinal blood vessels. Together the Endothelial cell of the retinal blood vessel and RPE is known as the Blood Retinal Barrier (BRB) because it regulates the diffusion of substances from the blood into the vitreous and retina⁸.

The blood vessels in the choroid have large fenestrations and it allows the diffusion of substances into and out of the choroidal stroma. The Bruch's membrane is membrane that separates the choroid from the RPE and it acts as the diffusional barrier and allows only macromolecules like proteins, oligonucleotides and genes. The RPE is the constituted of tight junctions which is the barrier to the entry of small molecules from the choroidal stroma to the retina¹³.

The efflux pumps like p-glycoproteins, multidrug resistance proteins (MRP) are observed on the RPE and it limits the permeation of various xenobiotics and endogenous compounds. Thus, both transcellular and paracellular passage of molecules across the RPE is restricted only to selected nutrients¹⁴.

In neural retina, the major blood vessels are in the innermost retinal layer which is the layer next to the inner limiting membrane which is the diffusional barrier between the retina and the vitreous. The innermost retinal layer has the ultra-structure of retinal blood capillaries which is uniform throughout the retina. These capillaries are of the endothelium are surrounded by the thick basement

membrane. Astrocytes are present close to the retinal blood vessels like blood vessels forming the blood brain barrier (BBB) in the brain¹⁵. The endothelium present is different. It does not show signs of fenestrations and has tight junction in a conserved manner. Thus, RPE and Retinal endothelial blood vessels collectively forms the BRB. It restricts the passage of molecules from the systemic circulation to the retina.

DISEASES ORIENTED WITH RETINA

The retina portion of the posterior segment of the eye is present at the back of eye and is responsible for providing images to the brain through the optic nerve. The retinal diseases and disorders alter the normal functioning of the eye which may lead to alteration of vision leading to the point of blindness. The common diseases include the floaters, macular degeneration, diabetic eye disease, retinal detachment and retinitis pigmentosa.

Floaters

Floaters is a condition in which a spot in the vision is observed. It may be age related but it also occurs in case of severe nearsightedness. In this condition, the jelly portion of the eye becomes more liquid and the clumps formed act as a shadow on the retina which is visualized as spots in vision^{9,16,17}.

Torn retina can result in floaters. In this case if the tear is not repaired the condition can turn into retinal detachment due to fluid accumulation behind the retina which is responsible for the separation from the eye¹⁸⁻²⁰.

Age-related Macular Degeneration (AMD)

AMD is the condition of the retina related to age causing central vision loss. In the individuals over the age of 55, this condition is commonly observed. Approximately 10 million individuals from the United States suffer from this condition. The symptoms are blurry central vision, difficulty in focusing fine details or warped straight lines. This condition can lead to blind spots. There are various treatments like antioxidant supplements which slow down the blocking of healthy blood vessel and progression of the condition.

Diabetic Macular Edema (DME)

For many patients suffering from diabetes, vision is also affected. The symptoms are blurry vision, floaters, double vision or dark spots, pressure or pain in at least one eye, flashing lights, trouble with peripheral vision or rings¹⁰.

DME is a condition where there is accumulation of fluid in the macula from leaking blood vessels. It is a manifestation of diabetic retinopathy, which causes damage to retinal vessels and vision impairment. Considering the rates of diabetes, the number of cases of diabetic retinopathy in the United States is expected to nearly double by 2050, rising from 7.7 million to 14.6 million. Approximately half of all people with diabetic retinopathy will develop DME²¹.

The treatment for diabetic eye disease is laser surgery. The patients suffering from diabetic macular edema are also at risk of cataracts and glaucoma.

Retinal Detachment

The retinal detachment occurs mainly if too much accumulation of fluid occurs behind the retina leading to separation of the retina. There are other factors that cause retinal detachment.

1. Genetic predisposition
2. Eye injury
3. Previous cataract surgery
4. Extreme near-sightedness
5. Previous retinal detachment in the other eye
6. The presence of other eye disorders

Floaters are indicators of the progress of retinal detachment. In some situations, flashes in the eye may occur and if not treated, it can lead to the permanent vision loss²². Other symptoms are seeing a gray curtain in the vision field or decrease in vision.

Retinitis Pigmentosa

It is genetic condition that causes retinal degradation. As the rods and cones die, the vision gradually declines^{23,24}. The conditions classified under retinitis pigmentosa are Usher syndrome, Leber's congenital amaurosis, rod-cone disease, refsum disease and Bardet-Biedel syndrome.

In general, the rods are firstly affected then followed by degeneration of the cones. The early symptom is night blindness but in some situations colour blindness or central vision loss occurs. Since it is an inherited condition, young adults and adolescents are especially vulnerable to this condition.

DRUG DELIVERY TO RETINA

The mode of drug administration affects the drug level attained in the posterior segment of the eye. The commonly employed modes of drug delivery to the retina are systemic

administration, topical administration, intravitreal injection local injections and ocular implants. Fig. 3 shows various routes of drug administration in retinal disorders.

The systemic drug delivery to the posterior segment of eye, mainly to the retina is difficult due to the presence of the BRB. Different strategies have been used for drug delivery to the retina²⁵⁻²⁷. An attempt has been made to increase transient permeability by modification of the barrier properties of RPE and the retinal endothelial cells by intracarotid infusion of hyper osmotic solutions like arabinose or mannitol. The osmotic gradient method is also used for nonspecific BRB opening but it leads to retinal and nervous system toxicity^{28,29}. Chemical modification to the drug molecule is also a widely used method to overcome the BRB. In this approach the drug is converted to lipophilic prodrug so that its transcellular diffusion into the neural retina is increased^{1,30}.

Systemic administration

The systemic delivery of drug for various ocular conditions has been attempted, but only 1 to 2% of plasma drug concentration has been achieved in the vitreous humor. Due to very low level of drug there is requirement of frequent administration of higher doses to achieve minimum therapeutic concentration. Treatment of acute human cytomegalovirus (HCMV) retinitis, includes foscarnet sodium or ganciclovir²³. The advantage of systemic administration is that it controls the spread of infection to other tissues but the disadvantage is nonspecific absorption by the other tissues may lead to serious toxicities. By considering the advantages and disadvantages of systemic administration for the treatment of ocular pathologies, it is not the ideal approach.

Topical route of administration

Topical applications of ophthalmic solutions and ointments are efficient drug delivery systems for the anterior segment of the eye (aqueous humor, iris ciliary body and lens), Table I lists topical formulation strategies to improve ocular bioavailability. For drug delivery to the posterior segment of the eye after topical administration, the drug has to traverse through the cornea, aqueous humor and lens. The precorneal constraints also affect bioavailability. Elimination of drug from the anterior segment occurs through the ciliary body or canal of Schlemm.

In the anterior chamber, the enzymatic metabolism that takes place that is responsible for loss of drug before reaching the posterior segment. Following topical administration, attaining the therapeutic drug level in the

posterior segment is difficult due to the presence of barriers. However according to reports, in the rabbit eyes therapeutic level of brimonidine (adrenergic agent) is achieved in the posterior segment after topical administration. This drug is used in glaucomatous neuropathy for retinal ganglion cell survival and neuroprotective action⁵⁶. According to literature upon topical administration therapeutic levels of brimonidine were achieved in choroid, retina and vitreous. The BRB penetration is confirmed by the vitreal drug level, which is relevant for neuroprotection⁵⁶.

The entry of verapamil in the posterior segment tissue after topical administration in rabbits is reported by Siegner *et al*⁵⁷. The drug concentration levels achieved were from lower to higher in serum, vitreous humor and aqueous humor respectively. But in treatment of infectious diseases such as endophthalmitis and CMV retinitis drug level does not reach minimum therapeutic level after topical administration.

Penetration studies of antibiotics from the systemic circulation to ocular fluids have confirmed that penetration to the vitreous is more restricted than penetration into the aqueous humor. Macha and Mitra studied aqueous and vitreous penetration of fluorescein (paracellular marker) in rabbits². They observed as compared to aqueous humor only 1 to 2% of plasma drug level of fluorescein is achieved in the vitreous humor. Attempts have been made to improve plasma drug level by increasing the amount of dose administered and rapid infusion but it resulted in increased ocular levels, however there is concern of long term systemic toxicity.

Intravitreal injections

Compared to the topical route, intravitreal administration offers certain advantages for drug delivery to the vitreous and retina. Intravitreal injection increases drug level in the posterior segment of the eye without causing systemic side effects. As this mode avoids BRB, therapeutic levels of drugs can be maintained with lower doses. In case of severe CMV retinitis and endophthalmitis intravitreal injection is the main therapy.

Intravitreal injection has become the backbone for treatment of posterior segment diseases over last 20 years. Antiviral drugs like acyclovir (ACV), ganciclovir (GCV) and foscarnet⁵⁸ cidofovir⁵⁹ and antibiotics like cefazolin, gentamicin⁷, cephalexin and ceftazidime⁶⁰ have been administered via the intravitreal route because it allows to optimize pharmacokinetic parameters so as to design proper dose and dosing frequency for the effective treatment of endophthalmitis and CMV retinitis⁶¹.

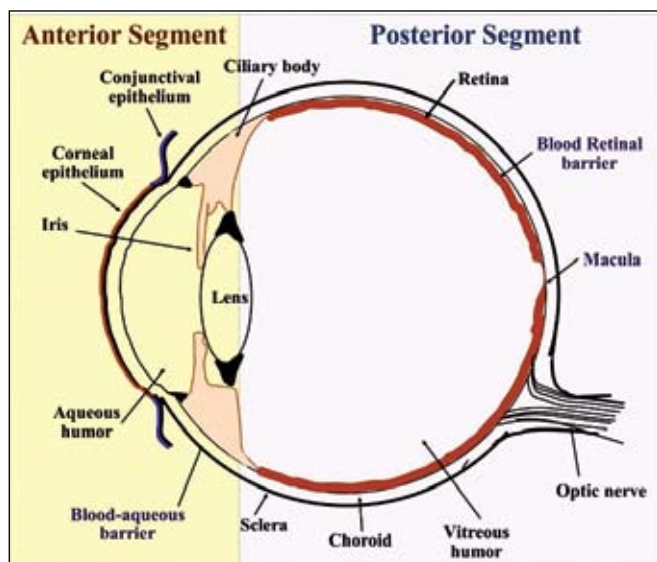


Fig. 1: Schematic Representation of anatomy of Eye

Intravitreal injection has major disadvantages like the need for repeated injections, patient noncompliance and infections associated with injections like endophthalmitis which can turn to severe conditions like retinal detachment. Researchers have attempted to overcome these shortcomings by increasing the residence time of the drug in vitreous after injection. The liposomes, lipophilic ester prodrugs and microparticles have been used. The particulate substance administration has the risk of blurred vision and risk of infections. The disposition of GCV and its monoester prodrugs have been studied by Macha and Mitra³⁰. They noticed that monoester prodrug of GCV generated sustained levels of drug in vitreous for longer periods of time compared to direct parenteral administration.

Intravitreal implants

Direct intraocular injections overcome the problem of high intravenous dose related toxicity. However, multiple injections are associated with an increased risk of cataract, astigmatism, endophthalmitis, retinal detachment and vitreous haemorrhage²⁴. Intravitreal implants have been developed as an alternative to multiple intravitreal injections. Vitrasert (Chiron Vision Corp., CA, USA) is a nonbiodegradable intraocular GCV implant. It is surgically inserted in the posterior segment of the eye, where it delivers GCV locally over a period of 5 – 8 months. Once the device is depleted of drug, it is replaced with a new device. Data from Chiron Vision Phase III trial demonstrated that disease progression was significantly delayed in patients implanted with Vitrasert compared to patients receiving intravenous doses of GCV⁶². However, the risks associated with intravitreal

injections are not completely eliminated. Adverse reactions reported for Vitrasert include loss of visual acuity, vitreous haemorrhage, retinal detachment, cataract formation, lens opacities, macular abnormalities, intraocular pressure spikes, optic disk/nerve changes, hyphemas, uveitis and acute or delayed onset endophthalmitis. Also, repeated surgeries are required for implantation and removal of the implant. Like intravitreal injections, intravitreal implants release drug locally into the vitreous humor and may not be able to stem the spread of infections to the contralateral eye. Thus, systemic administration is recommended in conjunction with intravitreal injections and implants.

Scleral drug delivery

The intravitreal injections and implants are associated with the risk of retinal detachment as result piercing for multiple injections and surgery. To overcome this issue, researchers developed scleral implants. It has been reported that sclera does not present a significant barrier for diffusion of drug into the vitreous. The sclera is found to be five times more permeable than the cornea. The passive diffusion is the primary mechanism of transscleral drug transport via the intracellular aqueous pore pathway. Rather than lipophilicity of the molecule, molecular size is the controlling factor in drug transport⁶³. There are two ways of scleral drug delivery to the posterior chamber of eye.

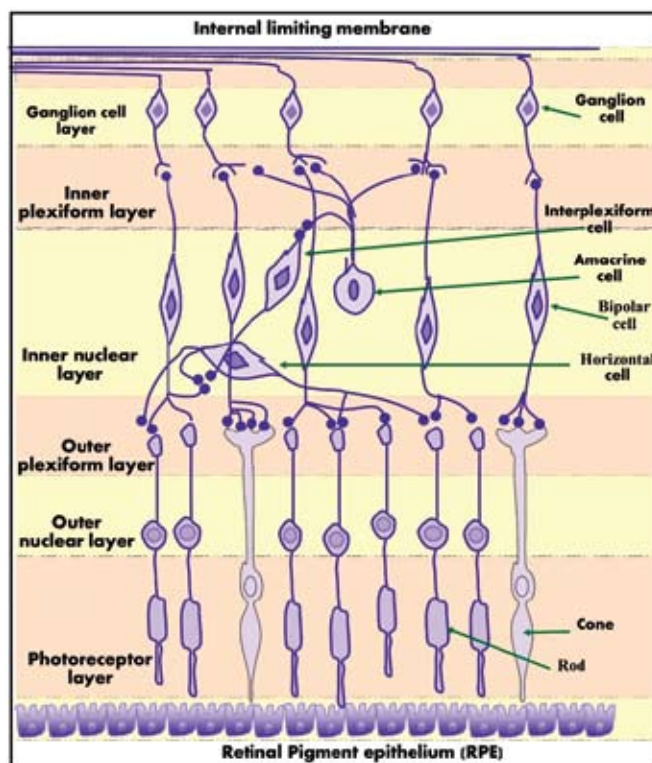


Fig. 2: Schematic Representation of Anatomy of Retina

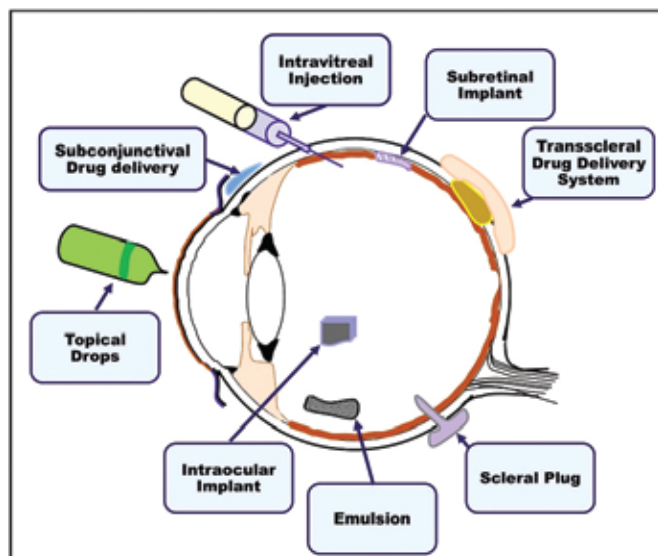


Fig. 3: Potential current drug delivery for retinal diseases

a) Scleral plugs and implants

Sakurai et al. developed biodegradable inserts of poly (DL-lactide) (PLA) and poly (DL-lactide-co-glycolide) (PLGA). Inserts were active in the HCMV inoculated eye for a period of 3 weeks. It reduced vitreo-retinal lesions compared to single intravitreal injection of GCV⁶³. In other study different blends of PLGA and PLA were examined for suitability in implant⁶⁴. The sustained GCV release over 24 weeks with therapeutic level above the minimum level without burst release effect for the entire period was obtained from PLA-70,000 and 5000 (80/20). The scleral implant can be used to achieve the sustained drug release. The disadvantages of such implants are patient discomfort due to the presence of a foreign body in the eye and the repeated operations involved⁶⁴.

b) Subconjunctival injections

The targeting of drugs to the posterior segment of the eye can be achieved by subconjunctival injections. The sclera and the conjunctiva are the primary surfaces for absorption. The absorption of timolol (β -blocker) upon corneal, scleral and conjunctival application into the ocular structure was studied by Sasaki *et. al.*⁶⁵. They observed higher plasma levels after conjunctival application. It can be concluded from the study that conjunctival absorption resulted in drug drainage into the systemic circulation. Scleral application resulted in increased drug level in the intraocular tissues as compared to conjunctival administration. The ocular pharmacokinetics of dacarbazine (anticancer drug) after subconjunctival and systemic administration was studied by Kalsi *et al.*⁶⁶. Upon subconjunctival administration elevated dacarbazine

levels were observed in the vitreous and aqueous humor compared with intravitreal administration.

Polymer based ophthalmic formulations were prepared to achieve sustained drug release of drug after subconjunctival drug injection. The biocompatible and biodegradable polymers are usually used for such purposes because polymers should not cause or induce inflammatory reactions in ocular tissues. The polymer should be degradable or erodible in a controlled manner to achieve zero-order release profile over a prolonged period. The polymers used in ocular drug delivery are pluronics, poloxamers, alginates, poly vinyl alcohol and polyorthoesters. The polymer's efficacy in providing drug release in subconjunctival administration is under investigation⁶⁷ biodegradable sustained release system for selected cases of glaucoma filtering surgery.

Transporters And Transporter-Mediated Drug Delivery

In spite of recent advances, drug delivery to the retina is still challenging because of several factors as this presence of MDR gene, lipophilicity and drug ionisation. The polar and hydrophobic properties of drugs (GCV, cidofovir, foscarnet) restrict their entry into retinal cell in conditions like CMV retinitis.

The entry into the cell by all active transport can be achieved by targeting the membrane transporters. This approach is successfully being used for improving the permeation of various molecules through biological membranes. In retinal drug delivery, after systemic, transscleral and intravitreal administration the intracellular drug concentration in retina can be increased by using this strategy.

A brief information on transporters and transporter mediated drug delivery and its applications in retinal drug delivery are discussed in following section.

Transporters

In transporter mediated drug delivery, the drug molecules are targeted to membrane transporters for the effective passage of the drug across the cell membrane. Different nutrient transporters like amino acid, folate, peptide, monocarboxylic acid, organic amino, nucleoside, nucleobase and cationic transporters are usually targeted⁶⁸. For targeting of parent drug to the membrane transporter, it is coupled with a promoiety which is an endogenous substrate for that transporter. The Promoiety

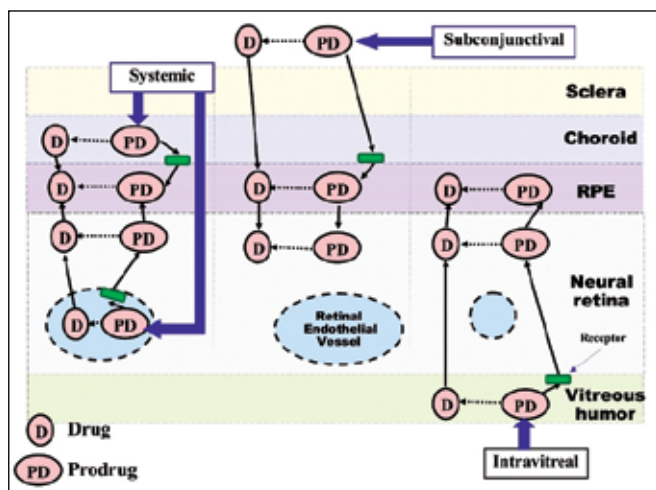


Fig. 4: Routes of Prodrug administration
(Adopted from Duvvuri S et. al.)

is a transporter which translocates the prodrug across the cell membrane. Then, the drug is released from the transporter intracellularly via enzymatic cleavage of the drug ligand bond. There is no toxicity issue from the ligand as it is a nutrient.

By using this approach, the transporters expressed on RPE and neural retina can be targeted with drug after systemic and intravitreal administration respectively (Fig. 4). As compared to conventional lipophilic prodrug design, transport targeted prodrug design has higher solubility and higher dose administration potential.

Different nutrient transporters expressed on the retina include glucose⁶⁹, peptide⁷⁰, GABA, taurine, proline, nucleoside-like adenosine⁷¹, glutamine, arginine, tryptophan and amino acids like glycine⁷². Among these transporters, those enhancing retinal drug delivery are discussed here.

a) Peptide transporter (PepTs)

These transports have been identified on epithelia. The information about their substrate specificity and mechanism is available⁷³⁻⁷⁵. For small molecules like di- and tripeptides, PepT1 and PepT2 have broad substrate specificity⁷⁶⁻⁷⁹. For easy transport across the lipophilic membrane the drug molecule is converted to a substrate known as prodrug modification⁷⁸. Due to this, the solubility of the substrate may be enhanced depending on type of amino acid selected in the peptide moiety. After the transport, the enzymes like dipeptidase, aminopeptidases and cholinesterases present in the retinal and vitreal tissue cleave the prodrug and then the a parent drug is regenerated. The molecules like zidovudine, GCV and ACV after converting into peptide prodrugs result

in a significant increase in the oral bioavailability of the parent drugs^{79,80}.

The studies confirm the presence of PepT on the RPE side facing blood and on the retina side facing the vitreous⁷⁰. The intravitreal administration of transporters facing the vitreous could be targeted to increase the intracellular drug concentration and after systemic administration transporters present on the RPE can increase vitreal and retinal drug concentrations.

b) Amino acid transporters

Amino acid transporters have been extensively studied on the blood brain barrier and the intestinal mucosa. These transporters are highly substrate specific and transport only specific amino acids. Different amino acid transporters such as glycine, tryptophan, glutamine, GABA and proline have been identified on the retina. The prodrugs can be designed for targeting the a carrier protein due to the expression of such a transporter system on the RPE or the retina⁷².

c) Monocarboxylic acid transport systems (MCTs)

The proton coupled MCTs take up and excrete pyruvate, monocarboxylic acids and lactate of mammalian cells⁸¹. The isoforms of MCT 1 to 9 have been identified. Monocarboxylic drugs transported through MCTs in the small intestine are pravastatin and salicylic acid⁸²⁻⁸⁴. The MCT transports carboxylate drugs and prodrugs after transscleral or systemic administration into the retina from the choroid or blood, respectively.

d) Folate transport system

For entry of folate into the mammalian cell, two transport processes have been reported. In one process transporter protein along with multiple membrane spanning domains known as reduced folate transporter is utilized. Instead of folate, protein interacts more effectively with the reduced folate⁸⁶. In the second process folate receptor binds and internalises bound folate through receptor mediated endocytosis. The liposome mediated drug delivery and the entry of anticancer drugs to cancer cells is facilitated by targeting them to folate receptors⁸⁷. The drug level in vitreous can be improved by targeting them to folate receptors. As in most normal tissues, folate receptor is absent except in the placenta and in the choroid plexus or, present at low levels. Drug delivery to the retina can be achieved after systemic administration by using a folate ester prodrug⁸⁸.

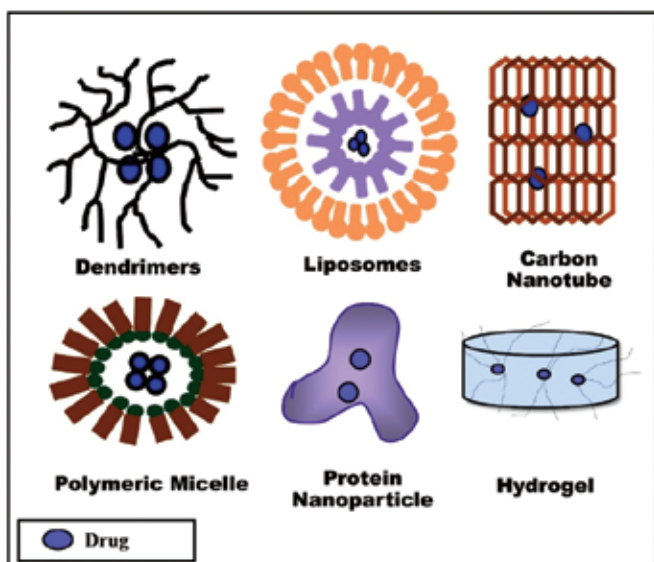


Fig. 5: Nanotechnological approaches for drug delivery (Adopted from Sibó Jiang *et al.*⁸⁹)

Strategies to improve ocular bioavailability by transporter-mediated drug delivery

Expression of transporters on the retina, particularly the RPE and endothelial cells of the retinal blood vessels, provides us with an opportunity to increase the retinal and vitreal levels of various drugs, thereby increasing their efficacy and decreasing the required dose. Compounds can be targeted to the transporter by delivery through any of the three main routes as follows¹.

a) Systemic administration of prodrugs

As discussed earlier, vitreal drug levels achieved upon systemic administration are around 1 – 2% of plasma levels. If a drug is targeted to a transporter expressed on the BRB, it could result in enhanced concentrations in the retina. Prodrug cleavage in the retinal cells may also facilitate greater drug levels in the vitreous humor.

The main disadvantage with this strategy is that drug uptake will be nonspecific, due to the possible distribution of the transporter at multiple tissues.

b) Intravitreal administration

In order to avoid the nonspecific absorption of drugs into all the tissues and to avoid systemic toxicity, intravitreal administration of prodrugs may be justified. In this strategy the prodrug is injected directly into the vitreous humor, thereby targeting it to the transporter expressed on the retinal cell membranes. Prodrug uptake

by the neural retina and RPE cells may be more efficient than the parent drug itself.

c) Subconjunctival administration

Subconjunctival administration usually results in higher vitreal concentrations than systemic administration. Moreover, subconjunctival injections of particulate dosage forms may result in sustained drug release over a prolonged period of time, thereby decreasing the frequency of administration. This route may be used to deliver prodrugs targeted to specific transporters expressed on the basolateral side of the RPE. Following subconjunctival administration, the prodrug first diffuses into the sclera and then into the choroidal circulation, where it interacts with transporters expressed on the RPE. These transporters will then carry the prodrug into the retinal tissue, where it is cleaved into the parent drug. If the drug is incorporated into a polymeric vehicle, which controls the release of the prodrug, a sustained delivery of the drug to the retina and vitreous humor may be achieved¹.

Nanotechnology in Retinal Drug Delivery

The Nanomaterials include nanosized particles with a size range of 1 to 100 nm. Different kinds of nanoparticles are used as tools in medicine⁸⁹. The nanoparticles have a higher surface area to volume ratio as compared with other macroscopic materials hence they have greater capability of carrying different drug molecules and facilitate greater attachment to the targeting moiety. Different nanoparticles with different size, shape and chemical characteristics can be engineered for a wide range of applications. The solubility of poorly water-soluble ophthalmic agents can be improved by attaching it to nanoparticles. Targeting the drug to the retina by enhancement of cellular drug uptake is a tool for drug transport through biological barriers. It increases the residence time and protects the drug from degradation. Because of the different advantages of the nanomedicines, it provides an excellent platform for designing the drug delivery system that is non-invasive and can deliver drug to retina in sustained manner⁸⁹.

The commonly employed nanoparticles include synthetic polymers (dendrimers, polymeric micelles, hydrogel), proteins (albumin nanoparticles), lipids (liposomes) and inorganic compounds (Cerium oxide nanoparticles)⁹⁰. Dendrimers are highly branched macromolecular structures while polymeric micelles are structures formed by self-assembly of amphiphilic copolymers with a hydrophobic core and a hydrophilic shell. Hydrogels are three-dimensional structures with

Table II: Nanoparticulate drug delivery to eye

Nanoparticle type	Drug	Route	Application	References
Pentablock copolymer	IgG-Fab fragment	General retinal disease	Intravitreal	99
Light responsive	Nintedanib	Neovascularization	Intravitreal	100
Liposome	Rpe65 DNA	Rpe65 associated blindness	Subretinal	101
PMMA dendrimer	Carboplatin	Retinoblastoma	Subconjunctival	43
PLGA	Bevacizumab	Wet AMD	Intravitreal	102
Naked siRNA	Claudin-5 siRNA	Modulation of BRB	Systemic	103

a high level of water content and having a network of hydrophilic polymers. The protein (like albumin) nanoparticles act by drug attachment to the protein surface. Albumin is a natural protein present abundantly in serum of most animals. Liposomes are the vesicles of lipid made up of lipid bilayer and enclose the aqueous core⁴⁷. Due to presence of a lipid bilayer and hydrophilic core, liposomes have the capability to carry both hydrophilic and lipophilic drugs^{91,92}.

To use nanotechnology in ocular drug delivery different properties of nanoparticles like size, shape and material surface properties need to be considered. The size of nanoparticles should be large enough so that it should not rapidly leak into the blood vessels and small enough to penetrate through the biological barrier of the retina⁹⁴. The study suggested that after intravitreal administration of 2 µm particles, particles remained in the vitreous fluid whereas for 200 nm sized particles, they uniformly distributed in the vitreous fluid and the inner limiting membrane of the retina. The 50 nm sized smaller nanoparticles were found in retina after two months of administration⁹⁵. The nanoparticles with similar size but different surface properties show different distribution inside the eye. Hence a retinal drug delivery system can be designed by changing the surface properties and optimizing the physicochemical properties of nanoparticles³⁵. For the drugs which cannot be delivered by other routes, their nanoparticles can be prepared for efficient delivery of such drugs. With the use of nanoparticles, there is a higher chance of drug reaching the target site at the back of the eye to treat the diseases affecting the retina. The clinical study for the safety and efficacy of nanoparticles are still in progress. Ideally the nanoparticles should be inert and biocompatible³⁴.

FDA has approved various polymers and excipients for human use like poly (ethylene glycol), poly lactic-co-glycolic acid (PLGA) and poly (hydroxyalkanoates) (PHAs). For ocular drug delivery chitosan, bovine serum albumin (BSA), poly acrylic acid (PAA) and Eudragit RS (ERS) have been used⁹⁶.

Despite the advantages of nanoparticles, there is a growing issue regarding potential toxicity of nanoparticles developed from the synthetic materials⁹⁷. Recently published literature on nanoparticles claims minimal or no toxicity of nanoparticles to cell lines and animals but very few literature focus modified of nanoparticles associated with toxicity⁹⁸. However, the toxicity of nanomaterials is to be examined in long term studies in future. Table II enlists various nanoparticulate drug delivery systems for the eye.

Fundamental Breakthroughs In Development Of Drug Delivery To Posterior Segment:

VITRASERT® (Ganciclovir insert)

Approval of the first sustained release posterior drug delivery system by FDA was in March 1996, 'Vitraser[®]'. Vitraser is a pellet of 4.5 mg ganciclovir coated with a biocompatible polymer by lamination for delivery of drug over long time for five to eight months⁶². This was for patients with acquired immunodeficiency syndrome (AIDS) for the local treatment of cytomegalovirus retinitis. The Vitraser[®] was withdrawn from the European market on 2nd April 2002 voluntarily by the marketing authorisation holder but the generic product may be available in market¹⁰⁴.

RETISERT® (fluocinolone acetonide intravitreal implant)

This intravitreal implant was approved by the FDA on 8th April 2005, for posterior segment chronic

Table III: Recent advances in Posterior segment drug delivery systems

Sr. No.	Advance Drug Delivery System	Manufacturer	Market Status	Reference
1	Yutiq™	Eye Point Pharmaceuticals	FdA Approved On 12 th October 2018; Commercially Available On 4 th February 2019	109
2	Xipere™	Clearside Biomedical	NDA Submitted To FDA On 19 th December 2018 Phase 3 Trials Complete	110
3	Port Delivery System	Roche/ Genentech	Sept. 2018 Phase 3 Trial Began	111
4	Gb-102	Graybug Vision	January 2019 Phase 1/2a Study Initial Data Analysis Reported Phase 2b Study Enrollment Expected In 2019	112
5	DexAmethasone Intravitreal Implant (Ar 1105) With Print® Technology	Aerie Pharmaceuticals	Ar-1105 Phase 2 Trial Began In Early 2019; Ar-13503 (Rho Kinase/Protein Kinase C Inhibitor) Phase 2 Trial To Be Initiated In June 2019	113

non-infectious uveitis. It is 0.59 mg sterile implant constructed to release fluocinolone acetonide locally to the posterior segment at an initial rate of 0.6 µg/day and decreasing after the first month to a steady state of 0.3-0.4 µg/day for approximately 30 months¹⁰⁵. It is a synthetic corticosteroid fluocinolone acetonide drug. This drug delivery system controls inflammation effectively but it requires surgery for its placement. It also has side effects such as increase in intraocular pressure (IOP) and cataract which require additional surgery for control¹⁰⁶.

Ozurdex® (Allergan)

It is a sustained release biodegradable intravitreal corticosteroid implant of 0.7 mg dexamethasone for six months treatment. On 17th June 2009, FDA approved this for the treatment of macular edema followed by retinal vein occlusion. Afterwards, it was approved for non-infectious uveitis of the posterior segment of the eye in 2010 and for diabetic the macular edema in 2014¹⁰⁷. Allergan voluntarily recalled 22 lots of this dosage form after noting detachment of approximately 300µm diameter of silicon particle from needle sleeve during administration¹⁰⁶.

Iluvien® (Alimera)

It is a non-bio erodible, sustained-release intravitreal implant approved by the FDA on 26th September 2014 for diabetic macular edema treatment. It is capable of continuous 36-month treatment of low dose corticosteroid after single injection¹⁰⁸.

Advances In Posterior Segment Drug Delivery Systems

Table III lists recent advances in posterior segment drug delivery. The table is followed by a detailed discussion.

YUTIQ™

YUTIQ™ (fluocinolone acetonide intravitreal implant) is indicated for the treatment of chronic non-infectious uveitis affecting the posterior segment of the eye. It is a non-bio erodible intravitreal implant containing 0.18 mg fluocinolone acetonide, designed to release fluocinolone acetonide at an initial rate of 0.25 mcg/day, and lasting 36 months¹⁰⁹.

XIPERE™

It is a suspension of corticosteroid triamcinolone acetonide for the treatment of macular edema associated with uveitis and administered through the back of the eye through suprachoroidal injection in the space between choroid and sclera. It is a proprietary treatment approach for rapid dispersion of drug at the back of the eye to reach adequate drug concentrations and stay at the a site of disease. It has potential to act for a longer period and minimizes harm to healthy parts. This approach provides the efficacy and requires less treatments¹¹⁰.

Port Delivery System (PDS)

Ranibizumab (PDS) is the port delivery system of Genentech. It is a small device which is refillable,

slightly longer than a rice grain. It is uniquely designed for continuous delivery of ranibizumab over time and surgically implanted in the eye.

In the standard treatment patient with Age Related Macular Degeneration (AMD), monthly eye injection PDS was developed to reduce frequent eye injections. It allows the people with wet AMD to refill the implant.

In the patients using PDS, it requires six months or a longer time for the first refill of the implant. In patients with a high dose, PDS has achieved the same vision outcome as ranibizumab eye injection¹¹¹.

GB-102

GB-102 is a depot formulation of sunitinib malate encapsulated in bioabsorbable microparticles intended for treatment of patients with wet AMD (leading cause of vision the loss in adult) via the intravitreal (IVT) route. It is next generation therapy for retinal diseases which has a drug-elution profile supporting twice a year IVT injections, hence, reducing the care associated with frequent injection. The sunitinib is a tyrosine kinase inhibitor (TKI) receptor blocker that potentially blocks the intracellular receptor which is known to be associated with proliferation, angiogenesis, fibrosis and vascular permeability. The sunitinib is also known to demonstrate neuroprotective effect to retinal ganglion cells and photoreceptors during preclinical models of neural injury²⁶. In this depot formulation, the microparticles aggregate into a biodegradable depot in the inferior vitreal cavity. The unique surface treatment of formulation minimizes the inflammations observed with traditional PLGA microparticles. Graybug is also planning development of GB-102 in other diseases of the retina including retinal vein occlusion (RVO) and diabetic macular edema (DME)¹¹².

Dexamethasone Intravitreal Implant (AR-1105) with PRINT® Technology

AR-1105 is biodegradable intravitreal PRINT® manufactured implant of steroid dexamethasone. The implant is expected to provide a six month duration of action and reduces multiple injections and care associated with other treatments of DME. Aerie filed an Investigational New drug (IND) application for AR-1105 in December 2018. The FDA has reviewed the IND and it is in effect. The Phase 2 Trial was initiated in March 2019 for treatment of macular edema due to retinal vein occlusion (RVO)¹¹³.

CONCLUSION

Drug delivery to the retina remains a challenge to all professionals treating retina ailments like Age related macular degeneration, Diabetic macular edema, Retinal pigmentosa, Retinal detachment and Floaters. In this review, we have attempted to give a comprehensive overview of treatment approaches developed by researchers to overcome challenges and the barriers associated with posterior segment especially retinal drug delivery with their advantages and disadvantages. This review has attempted to describe the magnitude of the challenges, efforts of researchers to accomplish safe and efficacious drug delivery to the retina.

REFERENCES

1. Duvvuri S, Majumdar S, Mitra AK. Drug delivery to the retina: challenges and opportunities. **Expert. Opin. Biol. Ther.**, 2003; 3:45–56.
2. Macha S, Mitra AK. Ocular Pharmacokinetics in Rabbits using a Novel Dual Probe Microdialysis Technique. **Exp. Eye Res.**, [Internet]. 2001 [cited 2019 Jul 5];72:289–99.
3. Sawaguchi S, Yue BYJT, Yeh P, Tso MOM. Effects of Intracameral Injection of Chondroitinase ABC In Vivo. **Arch. Ophthalmol.**, [Internet]. 1992 10:110.
4. Lundberg B, Behndig A. Separate and additive mydriatic effects of lidocaine hydrochloride, phenylephrine, and cyclopentolate after intracameral injection. **J. Cataract. Refract. Surg.**, 2008; 34:280–3.
5. Kim T, Hasan SA. A New Technique for Repairing Descemet Membrane Detachments Using Intracameral Gas Injection. **Arch. Ophthalmol.**, 2002;120:181.
6. Shah SG, Sridhar MS, Sangwan VS. Acute corneal hydrops treated by intracameral injection of perfluoropropane (C3F8) gas. **Am. J. Ophthalmol.**, 2005;139:368–70.
7. Ben-Nun J, Joyce DA, Cooper RL, Cringle SJ, Constable IJ. Pharmacokinetics of intravitreal injection. Assessment of a gentamicin model by ocular dialysis. **Invest. Ophthalmol. Vis. Sci.**, 1989;30:1055–61.
8. Cunha-Vaz JG. The blood-retinal barriers. **Doc Ophthalmol** 1976;41:287–327.
9. Murakami K, Jalkh AE, Avila MP, Trempe CL, Schepens CL. Vitreous Floaters. **Ophthalmol.**, 1983;90:1271–6.
10. Janoria KG, Gunda S, Boddur SH, Mitra AK. Novel approaches to retinal drug delivery. **Expert Opin Drug Deliv.** 2007; 4:371–88.
11. Shastri D, Shelat P, Shukla A, Patel P. Ophthalmic drug delivery system: Challenges and approaches. **Syst Rev Pharm.**, 2011;1:113.

12. Rizzolo LJ. Polarity and the development of the outer blood-retinal barrier. **Histol Histopathol.**, 1997;12:1057–67.
13. Marmor MF. Structure and function of the retinal pigment epithelium. **Int. Ophthalmol Clin.** 1975 [cited 2019 Jul 3];15:115–30.
14. Aukunuru JV, Sunkara G, Bandi N, Thoreson WB, Kompella UB. Expression of multidrug resistance-associated protein (MRP) in human retinal pigment epithelial cells and its interaction with BAPSG, a novel aldose reductase inhibitor. **Pharm. Res.**, 2001 [cited 2019 Jul 3]; 18:565–72.
15. Holash JA, Stewart PA. The relationship of astrocyte-like cells to the vessels that contribute to the blood-ocular barriers. **Brain Res.**, 1993 [cited 2019 Jul 3];629:218–24.
16. Stoffelns B, Vetter J, Keicher A, Mirshahi A. Pars Plana Vitrectomy for Visually Disturbing Vitreous Floaters in Pseudophacic Eyes. **Klin. Monbl. Augenheilkd.**, [Internet]. 2011 [cited 2019 Jul 3];228:293–7.
17. Dayan MR, Jayamanne DGR, Andrews RM, Griffiths PG. Flashes and floaters as predictors of vitreoretinal pathology: Is follow-up necessary for posterior vitreous detachment? **Eye** . 1996 .;10:456–8.
18. Glaser BM. Treatment of Giant Retinal Tears Combined with Proliferative Vitreoretinopathy. **Ophthalmol.**, 1986 [cited 2019 Jul 3];93:1193–7.
19. Glaser BM, Carter JB, Kuppermann BD, Michels RG. Perfluoro-octane in the Treatment of Giant Retinal Tears with Proliferative Vitreoretinopathy. **Ophthalmol.**, 1991 [cited 2019 Jul 3];98:1613–21.
20. Krishan NR, Chandra SR, Stevens TS. Diagnosis and Pathogenesis of Retinal Pigment Epithelial Tears. **Am. J. Ophthalmol.**, 1985;100:698–707.
21. Watch Out for Diabetic Retinopathy | Features | CDC [Internet]. [cited 2019 Jul 5].
22. Hollands H, Johnson D, Brox AC, Almeida D, Simel DL, Sharma S. Acute-Onset Floaters and Flashes. **JAMA.** 2009;302:2243.
23. Muccioli C, Belfort R. Treatment of cytomegalovirus retinitis with an intraocular sustained-release ganciclovir implant. **Brazilian J Med Biol Res = Rev Bras Pesqui medicas e Biol.** 2000;33:779–89.
24. Cantrill HL, Henry K, Melroe NH, Knobloch WH, Ramsay RC, Balfour HH. Treatment of cytomegalovirus retinitis with intravitreal ganciclovir. Long-term results. **Ophthalmol.**, 1989;96:367–74.
25. Gote V, Sikder S, Sicotte J, Pal D. Ocular Drug Delivery: Present Innovations and Future Challenges. **J. Pharmacol. Exp. Ther.**, 2019 [cited 2019 Jul 5]; jpet.119.256933.
26. Himawan E, Ekström P, Buzgo M, Gaillard P, Stefánsson E, Marigo V, et al. Drug delivery to retinal photoreceptors. **Drug Discov. Today**, 2019 [cited 2019 Jul 5];
27. Le Bourlais C, Acar L, Zia H, Sado PA, Needham T, Leverage R. Ophthalmic drug delivery systems - Recent advances. **Prog. Retin. Eye Res.**, 1998;17:33–58.
28. Millay RH, Klein ML, Shults WT, Dahlborg SA, Neuwelt EA. Maculopathy Associated With Combination Chemotherapy and Osmotic Opening of the Blood-Brain Barrier. **Am. J. Ophthalmol.**, 1986 ;102:626–32.
29. Frank JA, Dwyer AJ, Girton M, Knop RH, Sank VJ, Gansow OA, et al. Opening of blood-ocular barrier demonstrated by contrast-enhanced MR imaging. **J. Comput. Assist. Tomogr.**10:912–6.
30. Macha S. Ocular Disposition of Ganciclovir and Its Monoester Prodrugs following Intravitreal Administration Using Microdialysis. **Drug Metab. Dispos.** 2002; 30:670–5.
31. Gupta U, Agashe HB, Asthana A, Jain NK. A review of in vitro-in vivo investigations on dendrimers: the novel nanoscopic drug carriers. **Nanomed. Nanotech, Biol Med.** 2006;2:66–73.
32. Matricardi P, Di Meo C, Coviello T, Hennink WE, Alhaique F. Interpenetrating polymer networks polysaccharide hydrogels for drug delivery and tissue engineering. **Adv. Drug Deliv. Rev.**, 2013;65:1172–87.
33. Nanjawade BK, Manvi F V., Manjappa AS. In situ-forming hydrogels for sustained ophthalmic drug delivery. **J. Control. Rel.**, 2007;122:119–34.
34. Kumari A, Yadav SK, Yadav SC. Biodegradable polymeric nanoparticles based drug delivery systems. **Colloids Surfaces B Biointerfaces** 2010 [cited 2019 Jul 5]; 75:1–18.
35. Koo H, Moon H, Han H, Na JH, Huh MS, Park JH, et al. The movement of self-assembled amphiphilic polymeric nanoparticles in the vitreous and retina after intravitreal injection. **Biomaterials** 2012 ;33:3485–93.
36. Deshpande S, Sharma S, Koul V, Singh N. Core-shell nanoparticles as an efficient, sustained, and triggered drug-delivery system. **ACS Omega.** 2017;2: 6455–63.
37. Reis C, Neufeld R, Ribeiro A, Nanotechnology FV-:, and B, 2006 undefined. Nanoencapsulation I. Methods for preparation of drug-loaded polymeric nanoparticles. Elsevier [Internet]. [cited 2019 Mar 13];
38. Hsu KH, Gause S, Chauhan A. Review of ophthalmic drug delivery by contact lenses. **J. Drug Deliv. Sci. Technol.** 2014;24:123–35.
39. Hu X, Hao L, Wang H, Yang X, Zhang G, Wang G, et al. Hydrogel Contact Lens for Extended Delivery of Ophthalmic Drugs. **Int. J. Polym. Sci.**, 2011;2011:1–9.
40. Xinming L, Yingde C, Lloyd AW, Mikhalovsky S V., Sandeman SR, Howel CA, et al. Polymeric hydrogels

- for novel contact lens-based ophthalmic drug delivery systems: A review. **Contact Lens Anterior Eye.** 2008;31:57–64.
41. Maulvi FA, Soni TG, Shah DO. A review on therapeutic contact lenses for ocular drug delivery. **Drug Deliv.** 2016;23:3017–26.
 42. Ross AE, Bengani LC, Tulsan R, Maidana DE, Salvador-Culla B, Kobashi H, et al. Topical sustained drug delivery to the retina with a drug-eluting contact lens. **Biomaterials** 2019;119285.
 43. Shome D, Kalita D, Jain V, Sarin R, Maru G, Bellare J. Carboplatin loaded polymethylmethacrylate nano-particles in an adjunctive role in retinoblastoma: An animal trial. **Indian J. Ophthalmol.**, 2014;62:585.
 44. Li F, Wang T, He H, pharmaceuticals XT-I journal of, 2008 undefined. The properties of bufadienolides-loaded nano-emulsion and submicro-emulsion during lyophilization. Elsevier [Internet]. [cited 2018 Jan 19];
 45. Kaur IP, Garg A, Singla AK, Aggarwal D. Vesicular systems in ocular drug delivery: An overview. **Int J Pharm.** 2004;269:1–14.
 46. Mishra GP, Bagui M, Tamboli V, Mitra AK. Recent Applications of Liposomes in Ophthalmic Drug Delivery. **J. Drug Deliv.**, 2011;2011:1–14.
 47. Deshpande PP, Biswas S, Torchilin VP. Current trends in the use of liposomes for tumor targeting. **Nanomedicine** 2013 ;8:1509–28.
 48. Araújo J, Gonzalez E, Egea MA, Garcia ML, Souto EB. Nanomedicines for ocular NSAIDs: safety on drug delivery. **Nanomedicine Nanotechnology, Biol. Med.**, 2009;5:394–401.
 49. Mainardes R, Urban M, Cinto P, Khalil N, Chaud M, Evangelista R, et al. Colloidal Carriers for Ophthalmic Drug Delivery. **Curr. Drug Targets.** 2005;6:363–71.
 50. Tangri P, Khurana S. Basics of Ocular Drug Delivery Systems. **Int. J. Res. Pharm. Biomed. Sci.**, 2011;2:1541–52.
 51. Eljarrat-Binstock E, Domb AJ. Iontophoresis: A non-invasive ocular drug delivery. **J. Control. Rel.**, 2006;110: 479–89.
 52. Hosoya KI, Lee VHL, Kim KJ. Roles of the conjunctiva in ocular drug delivery: A review of conjunctival transport mechanisms and their regulation. **Eur. J. Pharm. Biopharm.**, 2005;60:227–40.
 53. del Amo EM, Urtti A. Current and future ophthalmic drug delivery systems. A shift to the posterior segment. **Drug Discov. Today.** 2008;13:135–43.
 54. Zderic V, Vaezy S, Martin RW, Clark JI. Ocular drug delivery using 20-kHz ultrasound. **Ultrasound Med Biol** [Internet]. 2002 [cited 2019 Jul 5];28:823–9.
 55. Sasaki H, Yamamura K, Mukai T, Nishida K, Nakamura J, Nakashima M, et al. Enhancement of Ocular Drug Penetration. **Crit. Rev. Ther. Drug Carr. Syst.** [Internet]. 1999 [cited 2019 Jul 5];16:62.
 56. Acheampong AA, Shackleton M, John B, Burke J, Wheeler L, Tang-Liu D. Distribution of Brimonidine into Anterior and Posterior Tissues of Monkey, Rabbit, and Rat Eyes. **Drug Metab. Dispos.**, 2002 ;30:421–9.
 57. SIEGNER SW, GIOVANONIRL, ERICKSONKA, NETLAND PA. Distribution of Verapamil and Norverapamil in the Eye and Systemic Circulation After Topical Administration of Verapamil in Rabbits. **J. Ocul. Pharmacol. Ther.**, 1998 ; 14:159–68.
 58. López-Cortés LF, Pastor-Ramos MT, Ruiz-Valderas R, Cordero E, Uceda-Montañés A, Claro-Cala CM, et al. Intravitreal pharmacokinetics and retinal concentrations of ganciclovir and foscarnet after intravitreal administration in rabbits. **Investig. Ophthalmol. Vis. Sci.**, 2001;42:1024–8.
 59. Cundy KC, Lynch G, Shaw J-P, Hitchcock MJM, Lee WA. Distribution and metabolism of intravitreal cidofovir and cyclic HPMPC in rabbits. **Curr. Eye Res.**, 1996 ;15:569–76.
 60. WAGA J, NILSSON-EHLE I, LJUNGBERG B, SKARIN A, STÄHLE L, EHINGER B. Microdialysis for Pharmacokinetic Studies of Ceftazidime in Rabbit Vitreous. **J. Ocul. Pharmacol. Ther.**, 1999 ;15:455–63.
 61. HUGHES PM, KRISHNAMOORTHY R, MITRA AK. Vitreous Disposition of Two Acycloguanosine Antivirals in the Albino and Pigmented Rabbit Models: A Novel Ocular Microdialysis Technique. **J. Ocul. Pharmacol. Ther.**, 1996; 12:209–24.
 62. Auritec Pharmaceuticals - Vitrsert® and Retisert® [Internet]. [cited 2019 Jul 1].
 63. Sakurai E, Matsuda Y, Ozeki H, Kunou N, Nakajima K, Ogura Y. Scleral Plug of Biodegradable Polymers Containing Ganciclovir for Experimental Cytomegalovirus Retinitis. **Invest. Ophthalmol. Vis. Sci.**, 2001;42:2043–8.
 64. UNLU N, ROBINSON JR. Scleral Permeability to Hydrocortisone and Mannitol in the Albino Rabbit Eye. **J. Ocul. Pharmacol. Ther.**, 1998;14:273–81.
 65. Sasaki H, Ichikawa M, Kawakami S, Yamamura K, Nishida K, Nakamura J. In Situ Ocular Absorption of Tilisolol Through Ocular Membranes in Albino Rabbits. **J. Pharm. Sci.**, 1996;85:940–3.
 66. Kalsi GS, Silver HK, Rootman J. Ocular pharmacokinetics of dacarbazine following subconjunctival versus intravenous administration in the rabbit. **Can. J. Ophthalmol.**, 199;26:247–51.
 67. Zignani M, Einmahl S, Baeyens V, Varesio E, Veuthey JL, Anderson J, et al. A poly(ortho ester) designed for combined

- ocular delivery of dexamethasone sodium phosphate and 5-fluorouracil: subconjunctival tolerance and in vitro release. **Eur. J Pharm. Biopharm.**, 2000 ;50:251–5.
68. Lee VH. Membrane transporters. **Eur. J. Pharm. Sci.** 2000;11 Suppl 2:S41-50.
 69. Ban Y, Rizzolo LJ. Regulation of glucose transporters during development of the retinal pigment epithelium. **Brain Res. Dev. Brain Res.**, 2000 ;121:89–95.
 70. Berger UV, Hediger MA. Distribution of peptide transporter PEPT2 mRNA in the rat nervous system. **Anat Embryol (Berl)** . 1999;199:439–49.
 71. Williams EF, Ezeonu I, Dutt K. Nucleoside transport sites in a cultured human retinal cell line established by SV-40 T antigen gene. **Curr Eye Res.** 1994 ;13:109–18.
 72. Pow D V. Amino acids and their transporters in the retina. **Neurochem Int.** 2001;38:463–84.
 73. Smith QR. Drug Delivery to Brain and the Role of Carrier-Mediated Transport. In: *Advances in experimental medicine and biology* 1993. p. 83–93.
 74. Chen L, Wright LR, Chen CH, Oliver SF, Wender PA, Mochly-Rosen D. Molecular transporters for peptides: delivery of a cardioprotective epsilonPKC agonist peptide into cells and intact ischemic heart using a transport system, R(7). **Chem Biol.** 2001;8:1123–9.
 75. Putnam WS, Pan L, Tsutsui K, Takahashi L, Benet L. Comparison of bidirectional cephalixin transport across MDCK and caco-2 cell monolayers: interactions with peptide transporters. - PubMed - NCBI. **Pharm Res.** . 2002.
 76. Kramer W, Dürckheimer W, Girbig F, Gutjahr U, Leipe I, Oekonomopoulos R. Influence of amino acid side-chain modification on the uptake system for β -lactam antibiotics and dipeptides from rabbit small intestine. **Biochim Biophys Acta - Biomembr.** 1990;1028:174–82.
 77. Meredith D, Temple CS, Guha N, Sword CJ, Boyd CAR, Collier ID, et al. Modified amino acids and peptides as substrates for the intestinal peptide transporter PepT1. **Eur J Biochem.** 2000;267:3723–8.
 78. Yang C, Tirucherai GS, Mitra AK. Prodrug based optimal drug delivery via membrane transporter/receptor. **Expert Opin Biol Ther.** 2001;1:159–75.
 79. Han H, de Vruet RL, Rhie JK, Covitz KM, Smith PL, Lee CP, et al. 5'-Amino acid esters of antiviral nucleosides, acyclovir, and AZT are absorbed by the intestinal PEPT1 peptide transporter. **Pharm Res.** 1998;15:1154–9.
 80. Pescovitz MD, Rabkin J, Merion RM, Paya C V, Pirsch J, Freeman RB, et al. Valganciclovir results in improved oral absorption of ganciclovir in liver transplant recipients. **Antimicrob Agents Chemother.** 2000;44:2811–5.
 81. Halestrap AP, Price NT. The proton-linked monocarboxylate transporter (MCT) family: structure, function and regulation. **Biochem J.** 1999;343 Pt 2:281–99.
 82. Takanaga H, Tamai I, Tsuji A. pH-dependent and carrier-mediated transport of salicylic acid across Caco-2 cells. **J Pharm Pharmacol.** 1994;46:567–70.
 83. Takanaga H, Maeda H, Yabuuchi H, Tamai I, Higashida H, Tsuji A. Nicotinic acid transport mediated by pH-dependent anion antiporter and proton cotransporter in rabbit intestinal brush-border membrane. **J. Pharm. Pharmacol** . 1996;48:1073–7.
 84. Hosoya K, Kondo T, Tomi M, Takanaga H, Ohtsuki S, Terasaki T. MCT1-Mediated Transport of L-Lactic Acid at the Inner Blood–Retinal Barrier: A Possible Route for Delivery of Monocarboxylic Acid Drugs to the Retina. **Pharm Res.** 2001;18:1669–76.
 85. Yoon H, Fanelli A, Grollman EF, Philp NJ. Identification of a Unique Monocarboxylate Transporter (MCT3) in Retinal Pigment Epithelium. **Biochem Biophys Res Commun.** 1997;234:90–4.
 86. Smith SB, Kekuda R, Gu X, Chancy C, Conway SJ, Ganapathy V. Expression of folate receptor alpha in the mammalian retinal pigmented epithelium and retina. **Invest Ophthalmol Vis Sci.** 1999;40:840–8.
 87. Anand P, Kunnumakkara AB, Newman RA, Aggarwal BB, Anand P, Kunnumakkara AB, et al. Bioavailability of Curcumin : Problems and Promises. 2007;4:807–18.
 88. Sudimack J, Lee RJ. Targeted drug delivery via the folate receptor. **Adv Drug Deliv Rev.** 2000;41:147–62.
 89. Duncan R, Gaspar R. Nanomedicine(s) under the Microscope. **Mol Pharm** 2011;8:2101–41.
 90. Cho K, Wang X, Nie S, Chen Z, Shin DM. Therapeutic Nanoparticles for Drug Delivery in Cancer. **Clin Cancer Res.** 2008;14:1310–6.
 91. Bozzuto G, Molinari A. Liposomes as nanomedical devices. **Int J Nanomed.** . 2015;10:975.
 92. Zhang R, He R, Qian J, Guo J, Xue K, Yuan Y. Treatment of Experimental Autoimmune Uveoretinitis with Intravitreal Injection of Tacrolimus (FK506) Encapsulated in Liposomes. **Investig Ophthalmology Vis Sci** 2010;51:3575.
 93. Jiang S, Franco YL, Zhou Y, Chen J. Nanotechnology in retinal drug delivery. **Int J Ophthalmol.** 2018;11:1038–44.
 94. Kim TW, Lindsey JD, Aihara M, Anthony TL, Weinreb RN. Intraocular distribution of 70-kDa dextran after subconjunctival injection in mice. **Invest Ophthalmol Vis Sci.** 2002;43:1809–16.
 95. Sakurai E, Ozeki H, Kunou N, Ogura Y. Effect of Particle Size of Polymeric Nanospheres on Intravitreal Kinetics. **Ophthalmic Res.** 2001;33:31–6.

96. Sahoo SK, Dilnawaz F, Krishnakumar S. Nanotechnology in ocular drug delivery. **Drug Discov Today**. 2008;13: 144–51.
97. El-Ansary A, Al-Daihan S, Bacha A Ben, Kotb M. Toxicity of Novel Nanosized Formulations Used in Medicine. In: Methods in molecular biology (Clifton, NJ). 2013. p. 47–74.
98. Heidel JD, Yu Z, Liu JY-C, Rele SM, Liang Y, Zeidan RK, et al. Administration in non-human primates of escalating intravenous doses of targeted nanoparticles containing ribonucleotide reductase subunit M2 siRNA. **Proc Natl Acad Sci**. 2007;104: 5715–21.
99. Agrahari V, Agrahari V, Hung W-T, Christenson LK, Mitra AK. Composite Nanoformulation Therapeutics for Long-Term Ocular Delivery of Macromolecules. **Mol Pharm**. 2016;13:2912–22.
100. Huu VAN, Luo J, Zhu J, Zhu J, Patel S, Boone A, et al. Light-responsive nanoparticle depot to control release of a small molecule angiogenesis inhibitor in the posterior segment of the eye. **J Control Rel.**, . 2015;200:71–7.
101. Rajala A, Wang Y, Zhu Y, Ranjo-Bishop M, Ma J-X, Mao C, et al. Nanoparticle-Assisted Targeted Delivery of Eye-Specific Genes to Eyes Significantly Improves the Vision of Blind Mice In Vivo. **Nano Lett**. 2014;14:5257–63.
102. Li F, Hurley B, Liu Y, Leonard B, Griffith M. Controlled Release of Bevacizumab Through Nanospheres for Extended Treatment of Age-Related Macular Degeneration. **Open Ophthalmol J**. 2012; 6:54–8.
103. Campbell M, Nguyen ATH, Kiang A-S, Tam LCS, Gobbo OL, Kerskens C, et al. An experimental platform for systemic drug delivery to the retina. **Proc Natl Acad Sci**. 2009;106:17817–22.
104. PUBLIC STATEMENT ON VITRASERT IMPLANT (Ganciclovir) . London; 2002.
105. RETISERT® Fluocinolone acetonide intravitreal implant) 0.59 mg Sterile 2007 [cited 2019 Jul 1].
106. Baker-Schena L. Drug Delivery for the Posterior Segment the Posterior Segment. *Eyenet Magazin* [Internet]. 2019;39–44.107. FDA approves expanded indication for Ozurdex implant. *FDA News* [Internet]. 2014;
108. Drugs@FDA: FDA Approved Drug Products (ILUVIEN) [Internet]. FDA. 2014 [cited 2019 Jul 1].
109. Home - Yutiq™ (fluocinolone acetonide intravitreal implant) 0.18 mg [Internet]. [cited 2019 Jul 1].
110. Clearside Biomedical Submits New Drug Application for XIPERE™ for the Treatment of Macular Edema Associated with Uveitis | Clearside Biomedical, Inc. - IR Site [Internet]. [cited 2019 Jul 1].
111. Genentech: Press Releases | Wednesday, Jul 25, 2018 [Internet]. [cited 2019 Jul 1].
112. GB-102 and GB-103 – Graybug Vision [Internet]. [cited 2019 Jul 1].
113. AR-1105 Implant - Aerie Pharmaceuticals Aerie Pharmaceuticals [Internet]. [cited 2019 Jul 1].



Indian Drug Manufacturers' Association (Event Calendar 2019-20)

Sr. No.	Day & Date	Organizer	Event	Venue
1.	Friday, 4 th October 2019	IDMA Contract Manufacturing Committee	Conference on “Creating Value Through Partnerships”	Rooftop & Malabar Hall, Hotel Trident, Nariman Point, Mumbai
2.	Saturday, 18 th January 2020	IDMA	“58th AGM & Annual Day Celebrations 2020”	Mumbai

For more details, please contact IDMA Secretariat at
 Email: admin@idmaindia.com / actadm@idmaindia.com, mail_idma@idmaindia.com or
 Mob: 9819035076/9821868758/9820629907 / Tel: 022-2494 4624 / 2497 4308