REVIEW ARTICLE

NANO-BASED THERAPY FOR TREATMENT OF SKIN CARCINOMA

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ABSTRACT

Skin carcinoma is a frequently occurring cancer caused due to ultra violet rays of the Sun. It starts from normal skin cells but later on transforms into cells which undergo uncontrolled mitosis. Skin cancer is not as deadly as other cancers and has no metastasis and is not life threatening. Conventional chemotherapy has in general failed to treat skin cancer due to non specific targeting, which is accompanied by several side effects. Novel therapeutic approach based on nanotechnology have emerged as the best alternative for skin cancer treatment. We presented current scenario of nano based particulate drug carrier approaches for effective therapy for skin carcinoma by reducing side effects. This approach also reduces frequency of administration and improves patient compliance. Nanotechnology has emerged as the best alternative for conventional therapy for the effective treatment of skin cancer. Nanoparticles can specifically target skin carcinoma and are able to sustain drug release and reduce side effects to a greater extent.

Keywords: Skin carcinoma, Nanoparticles, nanoemulsion, antineoplastics

INTRODUCTION

Nanotechnology is the branch of science which deals with formulations, substances as well as devices that run in nano sized range. It deals with design, fabrication and systems by minimizing size and shape in nano range^{1,2}. Nanotechnology is widely used in biomedical field such as imaging, diagnosis and therapy³. Due to site specific delivery potential of nanoparticles, they are widely used in the treatment of various types of cancers. Nanoparticles are also used for improving solubility of poorly water soluble drugs, bioavailability and modifying pharmacokinetic properties. Nanoparticles can enhance the biological half lives of drugs by decreasing metabolism. Combination therapy can be improved by using nanoparticles by simultaneous release of two or more drugs^{4,5}. Nanoparticles also reduce the dose and side effect as well⁶. The nanoparticles can be used for potentiating anticancer therapeutic effects by combining drug delivery with energy forms such as heat, light, and sound7. In the United States (US) skin cancer is widespread but not considered as deadly8.

Melanoma is a form of skin cancer which accounts for a majority of skin cancer deaths but this can be operated safely without harming the patient. The metastasized melanoma causes majority of deaths within short span of diagnosis i.e. within maximum of 5 years9. Apart from melanoma, other forms of skin cancers such as basal cell carcinoma and squamous cell carcinoma are frequently found types of skin cancer. Due to site specific drug delivery of nanoparticles they are used widely for improving therapeutic efficacy of anticancer drugs for skin cancer. The subsequent section deals with nanoparticles which are basically used both for systemic and transdermal delivery for effective treatment of skin cancers. The most important forms of nanoparticles which are employed for treatment of skin cancer include liposomes, dendrimers and carbon-based nanoparticles.

Liposomes

Liposomes are the concentric bilayers of phospholipids vesicles ranging from 50-100 nm and even larger enclosing a aqueous compartment. The classification of liposomes is based on the size and the number of layers. They may be classified as multi-, oligo- or unilameller. The aqueous layer provides the entrapment drugs which

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are soluble in water whereas the lipid double layers are used for retaining hydrophobic or amphiphilic drugs. To protect from the uptake of reticuloendothelial system (RES) after intravenous administration, PEGylated liposomes, stealth liposomes were designed which may be used for reducing clearance and enhancing circulation time¹⁰. The drug delivery specificity can be enhanced by linking the liposomal surface with suitable ligands and or polymers¹¹. Researchers found that liposomes reside in the interstitial medium of tumour cells¹². Several, so many liposomal formulations containing several anticancer drugs for treating melanoma are in the pipeline in clinical research¹³. Advancement in cationic liposomes led to targeting small interfering RNA (siRNA)¹⁴. Muthu and Feng developed theranostic liposomes which can be loaded with diagnostic NP¹⁵. Magnetic liposomes can be developed for tracing their path within the body using MRI¹⁶. Entrapment of drugs involved in ultrasound-controlled drug delivery systems can be done using magnetic liposomes¹⁷.

Solid Lipid Nanoparticles (SLNs)

SLNs are submicron colloidal drua delivery systems which are composed of physiological lipids dispersed in water or in aqueous surfactant solution. SLNs have no potential toxicity problems as organic solvents are not used during their preparation. SLNs are highly stable and can protect drugs against degradation. The other advantages of SLNs include biodegradability and biocompatibility and reportedly less toxicity. The production and sterilization of SLNs on commercial scale is also possible easily. Docetaxel loaded solid lipid nanoparticles (SLNs) was found to boost therapeutic efficacy of chemotherapeutic¹⁸ agents and malignant melanoma cell lines in vitro and in vivo studies¹⁹. Cholesteryl butyrate solid lipid nanoparticles were found effective for inhibiting HUVE (human umblilical vein endothelial cells adhesiveness to cancer) cell lines of different cancer types²⁰.

Nanospheres and polymeric micelles

They can be designed by 2 chains having variable hydrophobic tendency. They have both hydrophilic blocks and hydrophobic blocks²¹. The specificity and efficacy of micelles can be increased by modifying functional groups²²⁻²⁴. Polymeric micelle systems can also be employed for co-delivery of drugs along with radiation agents^{10,25,26}. B16F10 melanoma bearing mice were recently treated with polymeric micelles²⁷. Gadolinium (Gd) or manganese (Mn) are paramagnetic materials. They are generally developed as pH sensitive systems for oral delivery to protect the drugs from harsh conditions of the stomach²⁸.

Dendrimers

The term dendrimer is derived from a Greek word which means tree. Dendrimers are defined as molecules with repetitively branched units. Currently, the term dendrimer is internationally accepted. Dendrimers can be employed as carriers for delivering hydrophobic and hydrophilic drugs as well as, nucleic acids and imaging agents. Dendrimers possess excellent properties such as large number of surface functional groups, high number of branching and large number of internal cavities^{10,28-30}.

They include polyunsaturated fatty acids, oligo and polysaccharides and tumour associated antigens^{31–33}. Dendrimers have also successfully been employed for radio-immunotherapy and immunotherapy of different tumors and melanoma and squamous skin carcinoma^{34, 35}. Gadolinium-conjugate dendrimers have been employed for specific targeting as well as imagining of tumors³⁶.

Nanotubes

They are similar to graphite like carbon found in pencils. They are very thin hollow cylindrical tubes, much smaller than human hair but much stronger than steel and prepared either as single- or multi-walled nanotubes for use as tissue repair scaffolds and carrier of drugs³⁷.

SWCNT i.e. single-walled carbon nanotubes, are used by covalent bonding of large number of monoclonal antibodies for specific tumor targeting, fluorescent probes and radiation ion chelates³⁸. SWCNTs are used for carrying gadolinium atoms for MRI of tumors with specified receptor agonist and antagonist for targeting of tumor cells³⁹. The diagnosis and treatment of melanoma have been done by application of carbon nanotubes⁴⁰.

MSN (Mesoporous Silica Nanoparticles) It has come out a cost effective drug delivery system in the few last decades⁴¹⁻⁴³. MSNs are superior to conventional organic carriers due to their unique properties such as adjustable particle size, morphology, altered form, invariable and adjustable pore size, high surface area, high pore volume, high drug loading capacity and high mechanical as well as chemical stability and simple and easy functionalization⁴⁴⁻⁴⁶.

Quantum Dots Semiconductor crystals of nanometer dimensions (2-10 nm) with distinctive conductive properties determined by their size are known as Quantum dots. They have narrow emission band as well as broad absorption band⁴⁷. The middle part of quantum dots possess elements ranging from group II to VI of the periodic table⁴⁸, which are "over coated" with a layer of zinc sulphide. The photostability of quantum dots makes them



Fig. 1: Route of penetration of chemotherapeutic agents³⁴

suitable for exceptionally sensitive, durable and multitarget bio-imaging application^{49,50}. Peptides, antibodies and small molecules drugs are the possible ligands^{51.}

Novel approaches, such as a silica coating or a biocompatible polymer coating, have further increased the biocompatibility and reduced their toxicity.

When a magnetic field is applied externally, SPION (Superparamagnetic Iron Oxide) attains sufficient magnetic moment, for superparamagnetic behavior as they are used in biomedical fields.

In MRI, they are widely used as contrasting agents⁵². Very small quantities of SPIONs are needed for imaging therapy as they produce high contrast per unit of particles, thus able to reduce toxicity49,50. The SPIONs convert magnetic field into heat for selective destruction of tumor cells⁵³. This generated heat can selectively destroy the cancer cells which are more sensitive to temperature as compared to normal cells^{49,53}. Their biomedical applications can be enhanced by modifying their surface and increasing their biodegradability and biocompatibility⁵⁴. The attachment of biodegradable materials to SPION, such as dextran, PLGA, PEG and cellulose, can improve their biocompatibility and bioavailability. Recently nanoparticles coated with superparamagnetic iron oxide has been developed for sentinel lymph nodes mapping in breast cancer and melanoma patients⁵⁵.

AU Nanoparticles (Gold Nanoparticles) are used to carry drugs and genes. They are non-toxic carriers in which Au contributes stability to the system whereas monolayer imparts surface properties such as hydrophobicity and charge. Another most interesting characteristic of gold nanoparticles is their interaction with SH groups which provides an effective and selective means of controlled intracellular release. Recent developments in biochemical applications such as gene delivery, drug and imaging agents include formulation of gold nanoparticles like nanocapsules, nanotubes and nanospheres^{56,57}. The most attractive features of Au nanoparticles are excellent biocompatibility, ease of functionality, ease of preparation in nano range and their tendency to bind with different biomolecules with their original biological properties remain constant⁵⁸. Gold nanoparticles of size less than 50 nm can improve drug targeting to brain⁵⁹, sensitization of tissues and cells and in surgical processes⁶⁰⁻⁶². Drug targeting to tumour cells including melanoma have improved significantly by linking drugs, small molecules, proteins and DNA with gold nanoparticles. Gold nanoparticles have shown promising potential as biosensors as they can be detected by fluorescence, electrical conductivity and optical absorption^{63,64}. Moreover, they are nontoxic as well as biocompatible. Au nanoparticles are neither allergic nor immunogenic^{65,66}. Fig. 1 represents routes of penetration of chemotherapeutics for cure of skin carcinoma.

Nanoparticles as transdermal patches in skin carcinoma. Majority of the chemotherapeutic agents are commonly given parenterally. The topical application of anticancer drugs is found to be an alternative for drug targeting and therapeutic success⁶⁷. The major challenge of topical administration is to increase penetration of the antineoplastics drug to kill tumor cells⁶⁸. Penetration of drug through the major barrier of skin i.e. epidermis, can be improved by use of chemical permeation enhancers, sonophoresis, iontophoresis have been reported⁶⁹. Penetration of hydrophilic anticancer drugs through stratum corneum is low because of very less o/w partition coefficient value, significant molecular weight and ionization as well. Nanotechnology coupled with new imaging techniques have shown improved therapeutics benefits in skin cancer^{70,71}. The permeation of drug through stratum corneum is regulated

by Fick's 2nd law of diffusion⁷². 5-FU⁷³, diclofenac⁷³, and imiquimod are the currently used topical treatments for skin cancer as semisolid formulations. One of the most advanced and widely used treatment for skin USFDA is photodynamic therapy⁷⁴ which is used to treat non melanoma skin cancers and their precursor lesions, such as actinic keratosis. Nanoparticulate formulations could improve drug targeting and penetration into tumour cells, drug stability and reduce skin irritation⁷⁵. Liposomes are reported to be most widely used nanocarriers for treatment of skin cancer⁷⁶. Liposomes loaded with doxorubicin^{77, 78}, cisplatin^{79,80}, oxaliplatin⁸¹ and camptothecin⁸², have already proven improved cytotoxicity and minimized side effects. The liposomes, such as DOXIL, are already commercially available which contain doxorubicin and was approved in the US in 1995.

Fang *et al.*⁸³ reported that flexible liposomes (ethosomes) increased penetration through skin and retention in skin compared to traditional liposomes^{84,85} Similar results such as improved cytotoxicity and penetration was observed with 5-fluorouracil loaded niosomes⁸⁶. Liposome based formulations showed better therapeutic effect in the treatment of acne, psoriasis and other inflammatory conditions compared to non liposomes⁸⁷⁻⁹⁰; solid lipid nanoparticles and polymeric nanoparticles have shown increased drug penetration and stability^{91,92}.

DRUG DELIVERY FROM NANOPARTICLES IN NON MELANOMA SKIN CANCERS

There are basically three types of skin cancers i.e. melanoma, basal cell carcinoma (BCC), and squamous cell carcinoma (SCC). Among them, the incidence rate of BCC is 4 to 5 times higher than SCC⁹³. Exposure to ultraviolet radiation, skin type, family history, prior history of skin tumors, and immunosuppression are the primary reasons for sporadic skin malignancies. Excision is considered as the gold standard for the treatment of localized SSC and BBC. The various approaches available for excision are surgical excision, cryosurgery and micrographic surgery⁹⁴.

Majority of SCC and BCC are locally invasive, however 1 % to 5 % of primary SCC may permeate to nearby lymph nodes and distant locations as well⁹⁵. On the other hand, BCC can metastasize to distant sites of the body, which is considered a terminal condition⁹⁶. Topical treatment of SCC with 5-FU(5-fluorouracil) is widely used and recommended specially in situations where postoperative healing is impaired⁹⁷. There are many reactions for reoccurrence of tumour with typical treatment of 5-FU, such as incomplete duration of treatment, lower drug concentration in plasma, inadequate drug administration, poor penetration through epidermis⁹⁸. In order to improve penetration of 5-FU and to reduce many negative side effects of conventionally used chemotherapy drugs and control the release of the therapeutic agent, 5-FU loaded albumin magnetic nanocomposite spheres were reported to improve penetration and reduce side effects since albumin accumulates in tumor sites due to their altered physiology and metabolism. Administering polybutyl cyanoacrylate nanoparticles containing 5-FU once a day for 35 to 40 days for local remedy of patients with basal cell carcinoma showed absolute resolution illustrating that current practice is favoured by patients who are not surgical candidates⁹⁹. PDT (Photodynamic therapy), a nonsurgical treatment by application of a photo sensitizer (PS) in the presence of oxygen, can produce cytotoxic reactive oxygen species that oxidize subcellular organelles and biomolecules, ultimately leading to the destruction of diseased cells and tissues^{100,101}. Aminolevulinic acid, a PS agent using, blue light photodynamic therapy, is officially accepted for therapy, of actinic keratoses in Brazil, Argentina, Chile, USA, Korea, Mexico and Columbia. Methyl aminolevulinate, another PS agent, is licensed in Europe for PDT of AKs (actinic keratoses), Bowen's disease, and BCC¹⁰². Liposomes¹⁰³, chitosan¹⁰⁴ based nanoparticles and complexation with folic acid modified chitosan¹⁰⁵ was formulated for better penetration of ALA through epidermis. PLGA i.e. Poly (lactic-co-glycolic acid) based formulations¹⁰⁶ have shown encouraging results in the treatment of skin SCC A431 cells¹⁰⁷. For the prevention of skin carcinoma induced by UV radiation and BaP (benzo(a)pyrene) treatment in mice, Das et al. loaded Ap(apigenin), a dietary flavonoid having an anticancer property, with poly(lactic-coglycolide) nanoparticles (NAp)¹⁰⁸⁻¹⁰⁹. Antitumour activity by exposure to UV light has been reported in in vivo study in mice¹¹⁰. However, the nanoformulation of apigenin exhibited better effects than free apigenin and reduced tissue damage. Cytotoxic chemotherapy has not been approved for the treatment of advanced BCC. However, with variable successes, cisplatinum-based chemotherapy regimens have been used since 3 years¹¹¹. Recent advancements in understanding the pathogenesis of BCC lead to the development of targeting of drugs to the biological systems driving this carcinoma. Indeed, BCCs are critically dependent on a single signaling pathway, the sonic Shh (hedgehog) pathway, and the majority of BCC bearing mutations in genes in this developmental pathway¹¹², Sinces it has been illustrated that BCC tumor growth can be inhibited by blocking Shh-signaling¹¹³. Current literature reveals that nanoparticulate formulation of inhibitor of transcription factor (Gli 1, Nano HH1) has shown excellent tumour growth inhibition and antimetabolic effects¹¹⁴. Currently decarbazine (DTIC) is approved by FDA for first line treatment of melanoma¹¹⁵. As the median survival time of metastasized melanoma patients is around 6-10 months¹¹⁶⁻¹¹⁸, it has lead to the development of many particulate drug delivery systems such as liposomes¹¹⁹, dendrimers, polymersomes, carbon based nanoparticles and protein based nanoparticles^{120,121}. Gold nanoparticle of doxorubicin has shown improved chemotherapeutic effect beneficial against the melanoma cell lines¹²².

Lo Prete et al. prepared a nanoemulsion enriched with cholesterol to deliver etoposide to mice for the treatment of melanoma¹²³. They reported five times enhancement of maximum tolerated dose, significant decrease in adverse effects, and significant inhibition of growth of tumor by increasing the concentration of etoposide four times higher than the free etoposide at the tumor site. Nanoparticles loaded with doxorubicin conjugated with antibody against CD44, have shown significant drug targeting malignant cells¹²⁴. The size of the tumor is reduced to 60 % as compared to not treating tumor by the nanoparticles. For the treatment of metastatic melanoma, solvent-based taxanes were found to be active but demonstrated a high toxicity and limited efficacy due to poor solubility in water, resulting less loading as well as adverse effects to the solvents used in each formulation. Using (Nab-PTX) i.e. nanoparticles albumin bound paclitaxel, Phase II clinical trials as reported by Herch et al and Kottochade et al showed that albumin based paclitaxel nanoparticles were well tolerated in patients with metastatic melanoma. The response rate was 25.6 % in the chemotherapy-naive cohort and was 8.8% in the previously treated cohort¹²⁵. The inclusion of bevacizumab to nab-paclitaxel and carboplatin (regimen ABC) showed encouraging results in terms of both median progression-free survival and overall survival along with side effects vis-à-vis neutropenia, thrombocytopenia, neurosensory problems, fatigue, nausea and vomiting. In another clinical test¹²⁶, VEGF (vascular endothelial growth factor) antibody increased the effect of nab-PTX¹²⁷. Ott et al. found that the combination of a B cell lymphoma protein (Bcl)-2 antisense oligonucleotide, temozolomide, and nab-PTX produced a response of 40.6 %¹²⁸. Comparable side effects were also reported in Kottschade et al.'s study. Generally, higher cytotoxicity is observed with nanoparticles compared to free drug. Higher cytotoxicity was observed nearly 3.6 fold with phosphatidyl ethanolamine liposomal cisplatin compared to classical liposomes and free drug¹²⁹. Combination of inhibited melanoma proliferation and growth compared to monotherapy alone¹³⁰. Glucocorticoids loaded liposomes were highly effective in suppressing tumor angiogenesis as well as inflammation simultaneously¹³¹. Prednisolone phosphate loaded liposome was found to strongly inhibit endothelial cell proliferation and reduce proangiogenic protein (such as bFGF) levels, which were related to tumor angiogenesis¹³². Polyinosinic-polycytidylic acid loaded cationic liposome exceptionally enhanced tyrosinase related protein (TRP-2-specific) IFN-releasing cells which boosts the antitumor immune response. It opened another door for immunotherapy for melanoma by peritumoral injection¹³³. Fast build up as well as being contained within the tumor makes functionalized quantum dot-liposome hybrid a suitable approach^{134,135}. In vivo study of liposomal siRNA showed decreased melanoma growth and metastasis¹³⁶. Certain inhibitors of the MAPK pathway have been delivered by the most acceptable technology i.e. nanotechnology¹³⁷. Basu et al prepared nanoparticles entrapped with MEK1 inhibitor PD98059 and reported improved antitumour activity to cisplatinum. The above result has given a particular direction for research in tumor cell targeting. This led to the combination of BRAF inhibitors, MEK inhibitors with drug delivery systems to target brain metastasis. RNA interference based approach opens up the possibility of tumor-selective delivery of small RNA or DNA molecules targeting anticancer drugs to transcribed gene. A chitosan nanoparticle of VEGF siRNA demonstrated improved healing effect. A plasmid-based STAT3 siRNA delivered by functional graphene oxide which exhibited markedly minimized growth of xenografted tumor^{138,139}. In fact, STAT3 is regarded as one of the most important mediators in melanoma promoting brain metastasis¹⁴⁰. The increase efficacy of TNF-alpha (Tumor necrosis factor) was found by Inhibiting phosphorylated STAT3 for melanoma¹⁴¹. A nanoparticle can also carry siRNA against the oncogene c-Myc to target melanoma cells B16F10 and exhibited effectiveness against melanoma¹⁴². Increased anticancer effect was observed with nanoparticle formulation containing both siRNAs against BRAF and Akt3143. Recent research revealed that inclusion complexation of 4-hydroxynonenal with ß- cyclodextrin enhanced antitumor activity¹⁴⁴. Nanotechnology has been found to increase the therapeutic effect of Bcl-2 inhibition. In genetically modified i.e. mice without thymus, an antisense oligonucleotide against Bcl-2 i.e. oblimersen reduced the growth of xenografted melanoma¹⁴⁵. In phase III clinical trial in patients it was observed that the effectiveness of combination of oblimersen with DTIC

vascular-disruptive drug (combretastatin phosphate,

CA4P) and a liposomal formulation of doxorubicin) greatly

was twofold as compared to DTIC alone¹⁴⁶. Bcl-2 siRNA as well as Myc and VEGF loaded nanoparticle could be developed for the treatment of melanoma¹⁴⁷. It leads to Bcl-2 reduction in mRNA as well as levels of protein which enhanced the *in vitro* as well as *in vivo* anticancer activity. In phase I clinical trial, oblimersen in combination with temozolamide and nab-PTX found to be more effective in patients with advanced stage melanoma¹⁴⁸. Nanoformulations with immunotherapeutic agents have been found to reduce adverse effects^{148,149}.

Yao *et al.* developed nanoparticles with polyethylenimine linked to β – cyclodextrin and conjugated with folate, and again mixed with IL-2 plasmid. This combination reduced tumor growth as well as extended the survival of the mice with melanoma¹⁵⁰. Using poly (polycaprolactone), a biodegradable polymer, a nanoporous miniature device was developed for local delivery of cytokine IFN-alpha and exhibited constant slow release of IFN-alpha¹⁵¹.

CONCLUSION

Nanocarriers have emerged as the most promising and versatile capable system as they offer multidimensional benefits, like less irritation of skin and enhanced stability of entrapped drug. The key benefit of nanocarriers as drug delivery systems is increased penetration of anticancer drug through skin. The use of nano based drug delivery systems such as liposomes, polymeric nanoparticles and dendrimers have shown tremendous potential in enhancing drug permeation through skin, antitumor activity and site specificity.

REFERENCES

- 1. Singh R. and Lillard Jr. J.W.: Nanoparticle-based targeted drug delivery, **Exp. Molec. Pathol.**, 2009, 86(3), 215–223.
- Bharali D.J., Khalil M., Gurbuz M., Simone T.M. and Mousa S.A.: Nanoparticles and cancer therapy: a concise review with emphasis on dendrimers, Int. J. Nanomed., 2009, 4, 1–7.
- 3. Sanvicens N. and Marco M.P.: Multifunctional nanoparticles properties and prospects for their use in human medicine, **Trends Biotechnol.**, 2008, 26(8), 425–433.
- 4. Allen T.M. and Cullis P.R.: Drug Delivery Systems: Entering the Mainstream, **Science**, 2004, 303(5665), 1818–1822.
- Emerich D.F. and Thanos C.G.: The pinpoint promise of nanoparticle-based drug delivery and molecular diagnosis, **Biomol. Eng.**, 2006, 23(4), 171–184.
- 6. Jain R.K and Stylianopoulos T.: Delivering nanomedicine to solid tumors, **Nat. Rev. Clin. Oncol.**, 2010, 7(11), 653–664.
- Jabr-Milane L.S., Van Vlerken L.E., Yadav S. and Amiji M. M.: Multi-functional nanocarriers to overcome tumor drug resistance, Cancer Treat. Rev., 2008, 34(7), 592–602.

- Misak H., Zacharias N., Song Z., Hwang S., Man K.P., Asmatulu R. and Yang S.Y.: Skin cancer treatment by albumin/5-Fu loaded magnetic nanocomposite spheres in a mouse model, J. Biotechnol., 2013, 164(1), 130-136.
- 9. Jerant A.F., Johnson J.T., Sheridan C.D. and Caffrey T.J.: Early detection and treatment of skin cancer, **Am. Fam. Physician**., 2000, 62(2), 357-382.
- 10. Zhang L. and Zhang N.: How nanotechnology can enhance docetaxel therapy, **Int. J. Nanomed.**, 2013, 8, 2927–2941.
- 11. Torchilin V.P.: Recent advances with liposomes as pharmaceutical carriers, **Nat. Rev. Drug Discov.**, 2005, 4(2),145–160.
- Yuan F., Leunig M., Huang S.K., Berk D.A., Papahadjopoulos D. and Jain R.K.: Microvascular permeability and interstitial penetration of sterically stabilized (stealth) liposomes in a human tumor xenograft. **Cancer Res.**, 1994, 54(13), 3352–3356.
- Slingerland M., Guchelaar H. H. and Gelderblom H.: Liposomal drug formulations in cancer therapy: 15 years along the road. **Drug Discov. Today**, 2012, 17, 160–166.
- Yano J., Hirabayashi K., Nakagawa S., Yamaguchi T., Nogawa M., Kashimori I., Naito H., Kitagawa H., Ishiyama K., Ohgi T. and Irimura T.: Antitumor activity of small interfering RNA/cationic liposome complex in mouse models of cancer, Clin. Cancer Res., 2004, 10(22), 7721–7726.
- Muthu M.S. and Feng S.: Theranostic liposomes for cancer diagnosis and treatment: Current development and preclinical success. Expert Opin. Drug Deliv., 2013, 10(2), 151–155.
- Soenen S.J.H., Cocquyt J., Defour L., Saveyn P., Van der Meeren P. and de Cuype M.: Design development of magnetoliposome-based theranostics. Mater. Manuf. Proc., 2008, 23, 611-614.
- Huang S.: Liposomes in ultrasonic drug and gene delivery. Adv. Drug Deliv. Rev., 2008, 60(10), 1167-1176.
- Mehnert W. and M\u00e4der K.: Solid lipid nanoparticles: production, characterization and applications, Adv. Drug Deliv. Rev., 2001, 47(2-3), 165-196.
- Mosallaei N., Jaafari M.R., Hanafi-Bojd M.Y., Golmohammadzadeh S. and Malaekeh-Nikouei B.: Docetaxel-loaded solid lipid nanoparticles: preparation, characterization, *in vitro*, and *in vivo* evaluations. J. Pharm. Sci., 2013, 102(6), 1994-2004.
- 20. Minelli R., Serpe L. and Pettazzoni P. *et al.*: Cholesteryl butyrate solid lipid nanoparticles inhibit the adhesion and migration of colon cancer cells. **Br. J. Pharmacol.**, 2012,166(2), 587-601.
- 21. Torchilin V. P.: Micellar nanocarriers: pharmaceutical perspectives, **Pharm. Res.**, 2007, 24(1), 1-16.
- 22. Fonseca M.J., Jagtenberg J.C., Haisma H.J. and Storm G.: Liposome-mediated targeting of enzymes to cancer cells for site-specific activation of prodrugs: comparison with the corresponding antibody-enzyme conjugate, **Pharm. Res.**, 2003, 20, 423-428.

- Schnyder A., Krähenbühl S., Drewe J. and Huwyler J.: Targeting of daunomycin using biotinylated immune liposomes: pharmacokinetics, tissue distribution and *in vitro* pharmacological effects, J. Drug Target., 2005, 13(5), 325-335.
- 24. Omid C. Farokhzad, Jianjun C., Benjamin A. T. and Ines Sherifi, *et al.*: Targeted nanoparticle-aptamer bioconjugates for cancer chemotherapy *in vivo*. Proceedings of the **Proc. Natl. Acad. Sci. U S A**, 2006, 103(16), 6315-6320.
- Liangfang Z., Aleksandar F. R.M., Frank A., Frank X. Gu., Pamela A. B asto., Vaishali B., Sangyong J., Robert S L. and Omid C. F.: Co-delivery of hydrophobic and hydrophilic drugs from nanoparticle-aptamer bioconjugates, Chem. Med. Chem., 2007, 2(9), 1268-1271.
- Zhang L., Gu F.X., Chan J. M., Wang A. Z., Langer R. S. and Farokhzad O. C.: Nanoparticles in medicine: therapeutic applications and developments, Clin. Pharmacol. Ther., 2008, 83(5), 761-769.
- Coimbra M., F. Rijcken J. F., Stigter M., Hennink W.E., Storm G. and Schiffelers R.M.: Antitumor efficacy of dexamethasone-loaded core-cross linked polymeric micelles, J. Control. Release, 2012, 163(3), 361-367.
- Nazir S., Hussain T. A., Ayub A., Rashid U. and Macrobert A.J.: Nanomaterials in combating cancer: therapeutic applications and developments, Nanomedicine., 2013, 10(1), 19-34.
- Paleos C.M., Tsiourvas D., Sideratou Z. and Tziveleka L.: Drug delivery using multifunctional dendrimers and hyperbranched polymers, **Expert Opin. Drug Deliv.**, 2010, 7(12), 1387-1398.
- Svenson S.: Dendrimers as versatile platform in drug delivery applications, Eur. J. Pharm. Biopharm., 2009, 71(3), 445-462.
- Jaracz S., Chen J., Kuznetsova L.V. and Ojima I.: Recent advances in tumor-targeting anticancer drug conjugates, Bioorg. Med. Chem., 2005, 13(17), 5043-5054.
- Kojima C., Kono K., Maruyama K. and Takagishi T.: Synthesis of polyamidoamine dendrimers having poly(ethylene glycol) grafts and their ability to encapsulate anticancer drugs, **Bioconjug. Chem.**, 2000, 11(6), 910-917.
- Larocque J., Bharali D.J. and Mousa S.A.: Cancer detection and treatment: The role of nanomedicines, Mol. Biotechnol., 2009, 42(3), 358-366.
- 34. Chen J., Shao R., Zhang X.D. and Chen C.: Applications of nanotechnology for melanoma treatment, diagnosis, and theranostics, **Int. J. Nanomed.**, 2013, 8, 2677-2688.
- Battah S., Balaratnam S., Casas A., O'Neill S., Edwards C. and Alcira Batlle, *et al.* Macromolecular delivery of 5-aminolaevulinic acid for photodynamic therapy using dendrimer conjugates, **Mol. Cancer Ther.**, 2007, 6, 876-885.
- Histaka K. and Martin W. B.: Dendrimer-based macromolecular MRI contrast agents: characteristics and application. **Mol. Imaging**, 2003, 2(1), 1-10.
- 37. Polizu S., Savadogo O., Poulin P. and Yahia L.: Applications of carbon nanotubes-based biomaterials in biomedical

nanotechnology. **J. Nanosci. Nanotechnol**., 2006, 6(7), 1883-1904.

- Mc Devitt M.R., Chattopadhyay D., Kappel B.J., Jagg J.S. and Schiffman S. R., *et al.*: Tumor targeting with antibodyfunctionalized, radiolabeled carbon nanotubes, J. Nucl. Med., 2007, 48(7), 1180-1189.
- Bosi S., da Ros T., Spalluto G. and Prato M.: Fullerene derivatives: an attractive tool for biological applications, Eur. J. Med. Chem., 2003, 38(11-12), 913-923.
- 40. Naderi N., Madani S.Y., Ferguson E., Mosaheb A. and Seifalian A.M.: Carbon nanotubes in the diagnosis and treatment of malignant melanoma, **Anti. Cancer Agents Med. Chem.**, 2013,13(1), 171-185.
- 41. Keasberry N.A., Yapp C.W. and Idris A.: Mesoporous silica nanoparticles as a carrier platform for intracellular delivery of nucleic acids, **Biochem. Moscow**, 2017, 82, 655–662.
- Slowing I. I., Vivero-Escoto J. L., Wu C.W. and Lin V. S. Y.: Mesoporous silica nanoparticles as controlled release drug delivery and gene transfection carriers, Adv. Drug Deliv. Rev., 2008, 60(11), 1278-1288.
- 43. Torney F., Trewyn B.G., Lin V.S. and Wang K.: Mesoporous silica nanoparticles deliver DNA and chemicals into plants, **Nat. Nanotechnol.**, 2007, 2(5), 295-300.
- Slowing I., Trewyn B.G. and Lin V.S.Y.: Effect of surface functionalization of MCM-41-type mesoporous silica, J. Am. Chem. Soc., 2006, 128, 14792-14793.
- 45. Cauda V., Schlossbaue A., Kecht J., Zürner A. and Bein T.: Multiple core-shell functionalized colloidal mesoporous silica nanoparticles, **J. Am. Chem. Soc.**, 2009, 131, 11361-11370.
- Tsai C., Chen C., Hung Y., Chang F. C. and Mou C.: Monoclonal antibody-functionalized mesoporous silica nanoparticles (MSN) for selective targeting breast cancer cells, J. Mater. Chem., 2009, 19, 5737-5743.
- 47. Wang Y. and Chen L., Quantum dots, lighting up the research and development of nanomedicine, **Nanomedicine.**, 2011, 7(4), 385-402.
- 48. Alivisatos A.P.: Semiconductor clusters, nanocrystals, and quantum dots, **Science**, 1996, 271(5251), 933-937.
- Huang H., Barua S., Sharma G., Dey S.K. and Rege K. Inorganic nanoparticles for cancer imaging and therapy, J. Control. Release, 2011, 155(3), 344-357.
- 50. Medintz I. L., Uyeda H. T., Goldman E.R. and Mattoussi H: Quantum dot bioconjugates for imaging, labelling and sensing, **Nat. Mater.**, 2005, 4(6), 435-446.
- Ruoslahti E., Bhatia S.N. and Sailor M. J.: Targeting of drugs and nanoparticles to tumors, J. Cell Biol., 2010, 188, 759-768.
- 52. Ji T., Zhao Y., Ding Y. and Nie G.: Using functional nanomaterials to target and regulate the tumor microenvironment: Diagnostic and therapeutic applications, Adv. Mater., 2013, 25(26), 3508-3525.
- 53. Johannsen M., Thiesen, Burghard T., Peter W., Jordan and Andreas. *et al.*, Morbidity and quality of life during thermotherapy using magnetic nanoparticles in locally

recurrent prostate cancer: results of a prospective phase I trial, **Int. J. Hyperthermia**., 2007, 23(3) 315–323.

- 54. Kievit F.M. and Zhang M.: Surface engineering of iron oxide nanoparticles for targeted cancer therapy, **Acc. Chem. Res.**, 2011, 44(10), 853–862.
- Wang Y.X., Wang D.W., Zhu X.M., Zhao F. and Leung K.C.: Carbon coated superparamagnetic iron oxide nanoparticles for sentinel lymph nodes mapping, QIMS, 2012, 2(1), 53–56.
- Zavaleta C.L., Smith B.R., Walton I., Doering W, Davis G. and Shojaei *et al.*: Multiplexed imaging of surface enhanced Raman scattering nanotags in living mice using noninvasive Raman spectroscopy, Proceedings of the **PNAS USA.**, 2009, 106(32), 13511–13516.
- 57. Lu W., Ku G., Xiaoxia G., Wen, Zhou M., Guzatov D. *et al.*: Photoacoustic imaging of living mouse brain vasculature using hollow gold nanospheres, **Biomaterials**, 2010, 31(9), 2617–2626.
- 58. Albrecht R.: Immunocytochemistry: A Practical Approach, Oxford University Press, Oxford, UK 1993, PP. 244-248.
- Sonavane G., Tomoda K. and Makino K.: Biodistribution of colloidal gold nanoparticles after intravenous administration: effect of particle size, **Colloids Surf. B Biointerfaces.**, 2008, 66(2), 274-280.
- Dreaden E.C., Austin L.A., MacKey M.A. and El-Sayed M.A.: Size matters: Gold nanoparticles in targeted cancer drug delivery, **Ther. Deliv.**, 2012, 3(4), 457–478.
- Qian X., Peng X.H., Ansari D.O., Yin-Goen Q., Chen G.Z., Shin D.M., Yang L., Young A.N., Wang M.D. and Nie S.: *In vivo* tumor targeting and spectroscopic detection with surface-enhanced Raman nanoparticle tags, **Nat. Biotechnol.**, 2008, 26(1), 83–90.
- Jokerst J.V. and Gambhir S.S.: Molecular imaging with theranostic nanoparticles, Acc. Chem. Res., 2011, 44(10), 1050–1060.
- Huang X., Jain P.K., El-Sayed I.H. and El-Sayed M.A.: Gold nanoparticles: Interesting optical properties and recent applications in cancer diagnostics and therapy, Nanomedicine, 2007, 2(5), 681–693.
- Kimling J., Maier M., Okenve V., Kotaidis B. and Ballot H. A.: Plech A. Turkevich method for gold nanoparticle synthesis revisited, J. Phys. Chem. B., 2006, 110, 15700–15707.
- Hainfeld J.F., Dilmanian F.A., Slatkin D.N. and Smilowitz H.M. Radiotherapy enhancement with gold nanoparticles, J. Pharm. Pharmacol., 2008, 60(8), 977–985.
- Pan Y., Neuss S., Leifert A., Fischler M., Wen F. and Simon U. *et al.* Size-dependent cytotoxicity of gold nanoparticles, Small, 2007, 3(11), 1941–1949.
- Pan Y., Neuss S., Leifert A., Fischler M., Wen F., Simon U., Schmid G., Brandau W. and Jahnen-Dechent W.: Nanoparticles and microparticles for skin drug delivery, Adv. Drug Deliv. Rev., 2011, 63, 470–491.
- Delouise L. A.: Applications of nanotechnology in dermatology, J. Invest. Dermatol., 2012, 132, 964–975.
- 69. Taveira S.F. and Vianna Lopez R.F.: Topical administration of anticancer drugs for skin cancer treatment, in Topical

Administration of Anticancer Drugs for Skin Cancer Treatment. C. La Porta, Ed. pp. 247–272,

- Weiss M.B., Andrew E. and Aplin A.E.: Paying particle attention to novel melanoma treatment strategies, J. Investig. Dermatol, 2010, 130, 2699–2701.
- 71. Souza J.G., Gelfuso G.M., Simão P.S., Borges A.C. and Lopez R. F. V: Iontophoretic transport of zinc phthalocyanine tetrasulfonic acid as a tool to improve drug topical delivery. **Anti-Cancer Drugs**, 2011, 22(8), 783–793.
- 72. Williams A. C. and Barry B. W.: Penetration enhancers, Adv. Drug Deliv. Rev., 2004, 56(5), 603–618.
- Barrera M. V and Herrera E.: Topical chemotherapy for actinic keratosis and nonmelanoma skin cancer: current options and future perspectives. Actas. Dermosifiliogr., 2007, 98(8), 556–562.
- 74. Galiczynski E. M. and Vidimos A. T.: Nonsurgical treatment of nonmelanoma skin cancer, **Dermatol. Clin.**, 2011, 29(2), 297–309.
- 75. Schmid M. and Korting H. C.: Therapeutic progress with topical liposome drugs for skin disease, **Adv. Drug Deliv. Rev.**, 1996, 18, 335–342.
- 76. Gratieri T., Gelfuso G. M., Lopez R. F. and Souto E. B.: Current efforts and the potential of nanomedicine in treating fungal keratitis, **Expert Rev. Ophthalmol.**, 2010, 5(4), 365–384.
- 77. Hosoda J., Unezaki S., Maruyama K., Tsuchiya S. and Iwatsuru M.: Antitumor activity of doxorubicin encapsulated in poly(ethylene glycol)-coated liposomes, **Biol. Pharm. Bull.**, 1995, 18, 1234–1237.
- Barenholz Y.: Liposome application: problems and prospects, Curr. Opin. Colloid Interface Sci, 2001, 6(1), 66–77.
- 79. Chesnoy S. and Huang L: Structure and function of lipid-DNA complexes for gene delivery. **Annu. Rev. Biophys. Biomol. Struct.**, 2000, 29, 27-47.
- Krieger M.L., Schneider V., Koch M., Royer D., Jaehde U., Dieter H. and Eckstein N.: Overcoming cisplatin resistance of ovarian cancer cells by targeted liposomes *in vitro*, Int. J. Pharmaceutics, 2010, 389(1-2), 10–17.
- Abu Lila A. S., Doi Y., Nakamura K., Ishida T. and Kiwada H.: Sequential administration with oxaliplatin-containing PEG-coated cationic liposomes promotes a significant delivery of subsequent dose into murine solid tumor, J. Control. Release, 2010, 142, 167–173.
- Watanabe M., Kawano K., Toma K., Hattori Y. and Maitani Y.: *In vivo* antitumor activity of camptothecin incorporated in liposomes formulated with an artificial lipid and human serum albumin. J. Control. Release, 2011, 127, 231–238.
- Fang Y., Tsai Y., Wu P. and Huang Y.: Comparison of 5-aminolevulinic acid-encapsulated liposome versus ethosome for skin delivery for photodynamic therapy. Int. J. Pharmaceutics, 2008, 356, 144–152.
- Oh E. K., Jin S. E., Kim J. K., Park J., Park Y. and Kim C.: Retained topical delivery of 5-aminolevulinic acid using cationic ultra deformable liposomes for photodynamic therapy, **Eur. J. Pharm. Sci.**, 2011, 44, 149–157.

- Pierre M. B., Ricci Jr. E., Tedesco A. C. and Bentley M. V.: Oleic acid as optimizer of the skin delivery of 5-aminolevulinic acid in photodynamic therapy, **Pharm. Res.**, 2006, 23, 360–366.
- Paolino D., Cosco D., Muzzalupo R., Trapasso E., Picci N. Fresta M.: Innovative bola-surfactant niosomes as topical delivery systems of 5-fluorouracil for the treatment of skin cancer, Int. J. Pharm., 2008, 353, 233–242.
- Kitagawa S. and Kasamaki M.: Enhanced delivery of retinoic acid to skin by cationic liposomes. Chem. Pharm. Bull., 2006, 54, 242–244.
- Glavas-Dodov M., Fredro-Kumbaradzi E. and Goracinova K.: The effects of lyophilization on the stability of liposomes containing 5-FU, Int. J. Pharm., 2005, 291, 79–86.
- Rancan F., Papakostas D., Hadam S., Hackbarth S., Delair T. and Primard C. *et al.*: Investigation of polylactic acid (PLA) nanoparticles as drug delivery systems for local dermatotherapy, **Pharm. Res.**, 2009, 26, 2027–2036.
- Teixeira Z., Zanchetta B., Melo B.A., Oliveira L.L., Santana M.H., Paredes-Gamero E.J., Justo G.Z., Nader H.B., Guterres S.S. and Durán N.: Retinyl palmitate flexible polymeric nanocapsules: characterization and permeation studies, **Colloids Surf. B Biointerfaces**, 2010, 81(1), 374–380.
- Marquele-Oliveira F., Santana D.C., Taveira S.F., Vermeulen D.M., de Oliveira A.R., da Silva R.S. and Lopez R.F.: Development of nitrosyl ruthenium complex-loaded lipid carriers for topical administration: improvement in skin stability and in nitric oxide release by visible light irradiation, J. Pharm. Biomed. Anal., 2010, 53(4), 843–851.
- Misak H., Zacharias N., Song Z., Hwang S., Man K.P., Asmatulu R. and Yang S.Y.: Skin cancer treatment by albumin/5-Fu loaded magnetic nanocomposite spheres in a mouse model, J. Biotechnol., 2013, 164(1), 130–136.
- Joseph M. G., Zulueta W. P. and Kennedy P. J.: Squamous cell carcinoma of the skin of the trunk and limbs: the incidence of metastases and their outcome, **Aust. N. Z.** J. Surg, 1992, 62(9) 697–701.
- Wadhera A., Fazio M., Bricca G. and Stanton O.: Metastatic basal cell carcinoma: a case report and literature review. How accurate is our incidence data? **Dermatol. Online** J., 2006, 12(5), 5-7.
- 95. Sturm H. M.: Bowen's disease and 5-fluorouracil, J. Am. Acad. Dermatol., 1979, 1(6), 513–522.
- Mandekou-Lefaki I., Delli F., Koussidou-Eremondi T., Mourellou-Tsatsou O. and Dionyssopoulos A.: Imiquimod 5% cream: a new treatment for Bowen's disease, Int. J. Tissue React., 2005, 27(1), 31–38.
- 97. Goette D. K.: Topical chemotherapy with 5-fluorouracil, J. Am. Acad. Dermato., 1981, 4(6), 633–649.
- Hadjikirova M., Troyanova P. and Simeonova M.: Nanoparticles as drug carrier system of 5-fluorouracil in local treatment of patients with superficial basal cell carcinoma, J. BUON., 2005, 10(4), 517–521.
- 99. Huang Z., Xu H., Meyers A. D. and Lopez A.F.V.: Photodynamic therapy for treatment of solid tumors—

- 100. Morton C. A., Szeimies R. M., Sidoroff A. and Braathen L. R.: European guidelines for topical photodynamic therapy part 1: treatment delivery and current indications—actinic keratoses, Bowen's disease, basal cell carcinoma, J. Eur. Acad. Dermatol. Venereol., 2013, 27(5), 536–544.
- Morton C. A., McKenna K. E. and Rhodes L. E.: Guidelines for topical photodynamic therapy: update, **Br. J. Dermatol.**, 2008, 159(6), 1245–1266.
- 102. Casas A. and Batlle A.: Aminolevulinic acid derivatives and liposome delivery as strategies for improving 5-aminolevulinic acid- mediated photodynamic therapy, Curr. Med. Chem., 2006, 13(10), 1157–1168.
- 103. Yang V., Shieh M., Lin F. and Lou P.: Colorectal cancer cell detection by 5-aminolaevulinic acid-loaded chitosan nano-particles, **Cancer Lett**., 2009, 273(2), 210–220.
- 104. Yang S. J., Lin C. F., Kuo M. L. and Tan C. T.: Photodynamic detection of oral cancers with high-performance chitosanbased nanoparticles, **Biomacromolecules**, 2013, 14(9), 3183–3191.
- 105. Shi L., Wang X., Zhao F., Luan H, Tu Q, Huan Z. and Wang H.: *In vitro* evaluation of 5-aminolevulinic acid (ALA) loaded PLGA nanoparticles, **Int. J. Nanomedicine**, 2013, 8, 2669–2676.
- 106. Rejiya C. S., Kumar J., Raji V., Vibin M. and Abraham A.: Laserimmunotherapy with gold nanorods causes selective killing of tumour cells, **Pharmacol. Res.**, 2012, 65(2), 261–269.
- 107. Das S., Das J., Samadder A., Paul A. and Khuda-Bukhsh A. R.: Efficacy of PLGA-loaded apigenin nanoparticles in Benzo[a]pyrene and ultraviolet-B induced skin cancer of mice: Mitochondria mediated apoptotic signalling cascades, Food Chem. Toxicol., 2013, 62, 670–680.
- 108. Wei H., Tye L., Bresnick E. and Birt D. F.: Inhibitory effect of apigenin, a plant flavonoid, on epidermal ornithine decarboxylase and skin tumor promotion in mice, **Cancer Res.**, 1990, 50(3), 499–502.
- 110. Birt D. F., Mitchell D., Gold B., Pour P. and Pinch H. C.: Inhibition of ultraviolet light induced skill carcinogenesis in SKH-1 mice by apigenin, a plant flavonoid, **Anticancer Res.**, 1997, 17(1), 85–91.
- 111. Pfeiffer P., Hansen O. and Rose C.: Systemic cytotoxic therapy of basal cell carcinoma. A review of the literature, **Eur. J. Cancer**, 1990, 26(1), 73–77.
- 112. Hahn H., Wicking C., Zaphiropoulos P. G., Christiansen J., Chidambaram A. and Gerrard B.: A mammalian patched homolog is expressed in target tissues of sonic hedgehog and maps to a region associated with developmental abnormalities, **J. Biol. Chem.**, 1996, 271, 12125–12128.
- 113. Iwasaki J. K., Srivastava D., Moy R. L., Lin H. J. and Kouba D. J.: The molecular genetics underlying basal cell carcinoma pathogenesis and links to targeted therapeutics, J. Am. Aca. Dermatol., 2012, 66, 167–178.
- 114. Xu Y., Chenna V., Hu C., Sun H., Khan M. and Yang X.: Polymeric nanoparticle-encapsulated hedgehog pathway inhibitor HPI-1 (NanoHHI) inhibits systemic metastases in

an orthotopic model of human hepatocellular carcinoma, **Clin. Cancer Res.**, 2012, 18, 1291–1302.

- 115. Eigentler T. K., Caroli U. M., Radny P. and Garbe C.: Palliative therapy of disseminated malignant melanoma: a systematic review of randomised clinical trials, Lancet Oncol., 2003, 4, 748–759.
- 116. Falkson C. I., Ibrahim J., Kirkwood J. M., Coates A. S., Atkins M. B. and Blum R. H.: Phase III trial of dacarbazine versus dacarbazine with interferon α-2b versus dacarbazine with tamoxifen versus dacarbazine with interferon α-2b and tamoxifen in patients with metastatic malignant melanoma: an Eastern Cooperative oncology Group Study, J. Clin. Oncol., 1998, 16, 1743–1751.
- 117. Atkins M. B., Hsu J., Lee S., Cohen G.I., Flaherty L.E. and Sasman J.A.: Phase III trial comparing concurrent biochemotherapy with cisplatin, vinblastine, dacarbazine, interleukin-2, and interferon alfa-2b with cisplatin, vinblastine, and dacarbazine alone in patients with metastatic malignant melanoma (E3695): a trial coordinated by the Eastern Cooperative Oncology Group, **J. Clin. Oncol.**, 2008, 26(35), 5748–5754.
- 118. Sharma A., Sharma A. K., Madhunapantula S. V., Hue S. J., Desai D. and Amin S.: Targeting Akt3 signaling in malignant melanoma using isoselenocyanates, **Clin. Cancer Res.**, 2009, 15, 1674–1685.
- 119. Tran M. A., Watts R. J. and Robertson G. P.: Use of liposomes as drug delivery vehicles for treatment of melanoma, **Pigment Cell Melanoma Res.**, 2009, 22, 388–399.
- 120. Bei D., Meng J. and Youan B. C.: Engineering nanomedicines for improved melanoma therapy: progress and promises, **Nanomedicine**, 2010, 5, 1385–1399.
- 121. Pacheco I., Buzea C. and Tron V.: Towards new therapeutic approaches for malignant melanoma, **Expert Rev. Mol. Med.**, 2011, 13, 33.
- 122. Zhang X., Chibli H., Kong D. and Nadeau J.: Comparative cytotoxicity of gold-doxorubicin and InP-doxorubicin conjugates, **Nanotechnology**, 2012, 23(27), Article ID 275103.
- 123. lo Prete A. C., Maria D. A., Rodrigues D. G., Valduga C. J., Ibañez O. C. and Maranhão R. C.: Evaluation in melanomabearing mice of an etoposide derivative associated to a cholesterol-rich nanoemulsion. J. Pharm. Pharmacol., 2006, 58, 801–808.
- 124. Ndinguri M. W., Zheleznyak A., Lauer J. L., Anderson C. J. and Fields G. B.: Application of collagen-model triplehelical peptide-amphiphiles for CD44-targeted drug delivery systems, J. Drug Deliv., 2012, Article ID 59260.
- 125. Hersh E. M., O'Day S. J., Ribas A., Wolfram E. S. and Michael S. G.: A phase 2 clinical trial of nab-Paclitaxel in previously treated and chemotherapy-naive patients with metastatic melanoma, **Cancer**, 2010, 116,155–163.
- 126. Kottschade L. A., Suman V. J., Amatruda T., Amatruda A., McWilliams R. R. and Mattar B.I.: A phase II trial of nab-paclitaxel (ABI-007) and carboplatin in patients with unresectable stage IV melanoma, **Cancer**, 2011,117(8), 1704–1710.

- 127. Kottschade L. A., Suman V. J., Perez D. G., Vogt F.M., Hubner F. and Thorns C.: A randomized phase 2 study of temozolomide and bevacizumab or nab-paclitaxel, carboplatin, and bevacizumab in patients with unresectable stage IV melanoma: a north central cancer treatment group study, N0775, **Cancer**, 2013, 119, 586–592.
- 128. Ott P. A., Chang J. and Madden K.: Oblimersen in combination with temozolomide and albumin-bound paclitaxel in patients with advanced melanoma: a phase I trial, **Cancer Chemo. Pharmacol.**, 2013, 71(1), 183–191.
- 129. Hwang T., Lee W., Hua S. and Fang J., Cisplatin encapsulated in phosphatidylethanolamine liposomes enhances the *in vitro* cytotoxicity and *in vivo* intratumor drug accumulation against melanomas, **J. Dermatol. Sci.**, 2013, 46, 11–20.
- Mitrus I., Sochanik A., Cichoń T. and Szala S.: Combination of combretastatin A4 phosphate and doxorubicin-containing liposomes affects growth of B16-F10 tumors, Acta. Biochim. Pol., 2009, 56, 161–165.
- 131. Banciu M., Metselaar J. M., Schiffelers R. M. and Storm G.: Liposomal glucocorticoids as tumor-targeted antiangiogenic nanomedicine in B16 melanoma-bearing mice, J. Steroid Biochem. Mol. Biol., 2008, 111, 101–110.
- 132. Banciu M., Schiffelers R. M., Fens M. H. A. M., Metselaar J. M. and Storm G.: Anti-angiogenic effects of liposomal prednisolone phosphate on B16 melanoma in mice, J. Control. Release, 2006,113, 1–8.
- 133. Tran M. A., Watts R. J. and Robertson G. P.: Use of liposomes as drug delivery vehicles for treatment of melanoma. **Pigment Cell Melanoma Res.**, 2009, 22(4), 388–399.
- 134. Fujimura T., Nakagawa S., T. Ohtani, Ito Y. and Aiba S.: Inhibitory effect of the polyinosinic-polycytidylic acid/cationic liposome on the progression of murine B16F10 melanoma. **Eur. J. Immunol.**, 2006, 36, 3371–3380.
- 135. Al-Jamal V., Al-Jamal K.T., Bomans P.H., Frederik P.M. and Kostarelos K.: Functionalized-quantum-dot-liposome hybrids as multimodal nanopartides for cancer, **Small**, 2008, 4, 1406–1415.
- 136. Villares G. J., Zigler M., Wang H. and Melnicova V.: Targeting melanoma growth and metastasis with systemic delivery of liposome-incorporated protease-activated receptor-1 small interfering RNA, Cancer Res., 2008, 68, 9078–9086.
- 137. Inamdar G. S., Madhunapantula S. V. and Robertson G. P.: Targeting the MAPK pathway in melanoma: why some approaches succeed and other fail, **Biochem. Pharmacol.**, 2010, 80, 624–637.
- 138. Basu S., Harfouche R., Soni S., Chimote G., Mashelkar R. A. and Sengupta S.: Nanoparticle-mediated targeting of MAPK signaling predisposes tumor to chemotherapy, **PNAS USA**, 2009, 106, 7957–7961.
- 139. Yin D., Li Y., Lin H. and Guo B.: Functional graphene oxide as a plasmid-based Stat3 siRNA carrier inhibits mouse malignant melanoma growth *in vivo*, **Nanotechnology**, 2013, 24, 10, Article ID 105102.

- 140. Xie T., Huang F., Aldape K. D., Liu M., Gershenwald J.E. and Sawaya R. *et al.*: Activation of Stat3 in human melanoma promotes brain metastasis, **Cancer Res**., 2006, 66, 3188–3196.
- 141. Kong L., Gelbard A., Wei J., Reina-Ortiz C., Wang Y. and Yang E. C.: Inhibition of p-STAT3 enhances IFN- α efficacy against metastatic melanoma in a murine model, **Clin. Cancer Res.**, 2010, 16, 2550–2561.
- 142. Chen Y., Bathula S. R., Yang Q. and Huang L.: Targeted nanoparticles deliver siRNA to melanoma, **J. Investigative Dermatol.**, 2010, 130, 2790–2798.
- 143. Tran M. A., Gowda R., Sharma A. and Park E.J.: Targeting V600EB-Raf and Akt3 using nanoliposomal-small interfering RNA inhibits cutaneous melanocytic lesion development, **Cancer Res.**, 2008, 68, 7638–7649.
- 144. Pizzimenti S., Ciamporcero E., Pettazzoni P., Katiyar S.K., Ballestas M.E. and Athar M. *et al.*: The inclusion complex of 4-hydroxynonenal with a polymeric derivative of β-cyclodextrin enhances the antitumoral efficacy of the aldehyde in several tumor cell lines and in a three-dimensional human melanoma model, Free Radical Biol. Medicine, 2013, 65, 765–777.
- 145. Benimetskaya L., Ayyanar K., Kornblum N., Castanotto D., Rossi J. and Wu S.: Bcl-2 protein in 518A2 melanoma cells *in vivo* and *in vitro*. **Clin. Cancer Res.**, 2006, 12, 4940–4948.

- 146. Bedikian A. Y., Millward M., Pehamberger H, Rudin C. M., Salgia R. and Wang X.: Bcl-2 antisense (oblimersen sodium) plus dacarbazine in patients with advanced melanoma: the oblimersen melanoma study group, J. Clin. Oncol., 2006, 24, 4738–4745.
- 147. Beloor J., Choi C. S., Nam H. Y. and Kumar P.: Arginineengrafted biodegradable polymer for the systemic delivery of therapeutic siRNA, **Biomaterials**, 2012, 33, 1640–1650.
- 148. Zhang Z., Tongchusak S., Mizukami Y., Joong K.Y. and Tetsua L.: Induction of anti-tumor cytotoxic T cell responses through PLGA-nanoparticle mediated antigen delivery, **Biomaterials**, 2011, 32, 3666–3678.
- 149. Perche F., Benvegnu T., Berchel M., Lebegue L., Pichon C. and Jaffres P.A.: Enhancement of dendritic cells transfection *in vivo* and of vaccination against B16F10 melanoma with mannosylated histidylated lipopolyplexes loaded with tumor antigen messenger RNA, **Nanomedicine**, 2011, 7(4), 445–453.
- 150. Yao H., Ng S. S., Huo L., Shen Z, Yang M. and Li M.: Effective melanoma immunotherapy with interleukin-2 delivered by a novel polymeric nanoparticle, **Mol. Cancer Ther**., 2011, 10, 1082–1092.
- 151. He H., Grignol V. and Karpa V.: Use of a nanoporous biodegradable miniature device to regulate cytokine release for cancer treatment. **J. Control. Release**, 2011, 151, 239–245.



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