

REVIEW ARTICLE

VERSATILE APPROACHES FOR ANALYTICAL METHOD VALIDATION OF ANTICANCER DRUGS: A REVIEW

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ABSTRACT

Cancer refers to a group of illnesses that result from cell population in the body increasing unusually. These cells break up and create new cells in an uninhibited mode that can extend in the body and cause injury to vital organs. Analytical chemistry is the division of chemistry involved in separating, identifying and determining the relative quantity of the components in a sample. Analytical method development and validation play vital role in method development and manufacture of pharmaceuticals. The objective of this review article is to study divergent types of anticancer drugs and the different analytical methods assessed during their determination, like UV-Visible Spectrophotometer, GC, Mass Spectrophotometer, NMR, LC-MS, GC-MS and FT-IR. The involvement for analytical methods to establish an anticancer drug is of utmost importance. The development and validation of analytical methods is mandatory for preclinical and clinical studies and even for the development of formulations containing these compounds. This constitutes the next challenge in the analysis of anticancer drugs. This review outlines the recent position of method development and validation of anticancer drugs in bulk and solid dosage forms.

Keywords: Anticancer drug, method development and validation, ICH guidelines, UV Visible spectroscopy, RP-HPLC, LC-MS

INTRODUCTION

Analytical instruments play a major role in method development to get high quality and consistent analytical data. Analytical method development is the process of selecting a precise method to establish the composition of a formulation¹. Analytical method could be spectral, chromatographic, electrochemical or hyphenated. Method development and validation are continuous and mutually supporting tasks connected with research and development, quality control and quality assurance departments^{2,3}. Analytical procedures play a decisive role in equivalence and risk assessment and their management.

Cancer is a disease in which the control of growth is missing in one or more cells, leading to formation of a solid dosage mass of cells known as a tumour or to a blood or bone marrow linked cancer⁴. These cells divide and create new cells in an uninhibited manner that can increase in the body and cause injury to vital organs. Anticancer drugs are grouped according to the therapy as chemotherapy, hormonal therapy, and immunotherapy. Chemotherapeutic drugs include a family defined equally by their chemical structure and mechanism of action: alkylating agents, antibiotics, antimetabolites, topoisomers, primary and secondary inhibitors, mitosis inhibitors and others^{5,6}.

Alkylating agents: Alkylating agents have been used for the treatment of cancer for more than six decades⁷. These agent acts for the duration of all stages of the cell cycle, directly on DNA, cross linking the N-7- guanine

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residues, cause DNA filament breaks, leading to irregular base combination, disruption of cell distribution and finally resulting in cell death^{8,10}. Analytical methods for determination of alkylating agents are summarized in Table I.

Platinum coordination complexes: The platinum coordination complexes are a category of anticancer agents that are typically classified as alkylating agents, but which are of individual types¹². Their anticancer activity appears to transmit to the fractuous connecting of DNA molecules in a manner analogous to alkylating

agent¹⁴. Analytical methods for determination of platinum coordination complexes are summarised in Table II.

Antimetabolites: Antimetabolites are substances which, by virtue of their comparable chemical structure to vital compounds in the cell, interfere with the consumption of these substances and therefore harmfully affect cell utility and enlargement. Antimetabolites are frequently term “analogues,” and different types are well known¹⁶. Analytical methods for determination of antimetabolites are given in Table III.

Table I: Analytical methods for alkylating agents

Sr. No.	Sub-Class	Drug name	Instrument used for method development	Solvent system	Reference
1.	Nitrogen mustard	Cyclophosphamide	RP-HPLC	A-Water-acetonitrile (9:1V/V) B-Water-acetonitrile (3:7 V/V)	8
			LC-MS/MS	Triple liq.liq. extraction-ethyl acetate-dichloromethane (3:1:6 V/V/V)	9
			SPE-HPLC-MS/MS	Methanol-water (1:1 V/V)	10
			HPLC-UV	0.05M Disodium hydrogen phosphate-acetonitrile (65:35 V/V)	11
			UPLC	Acetonitrile-pH-7.0 buffer (20:80 V/V)	12
			LC	Acetonitrile-water (60:40V/V)	13
			GC-MS	Diethyl ether-ethyl acetate	14
			UV-VIS-Spectrophotometer	0.1N HCl	15
		Ifosphamide	UPLC-MS/MS	Ammonium formate-methanol-acetonitrile (40:48:12 V/V/V)	16
			RP-HPLC	Methanol	17
			SPE-HPLC-UV	Phosphate buffer pH-6.0- acetonitrile (77.25:22.75 V/V/V)	18
		Chlorambucil	UV-Visible-Spectrophotometer	Acetonitrile	19
			LCMS/MS	Methanol	20
		Bendamustine HCl	RP-HPLC	0.1Trifluoroacetic acid in water- acetonitrile (90:10 V/V)	21
			RP-HPLC	Trifluoroacetic acid and acetonitrile	22

			UV-VIS-Spectrophotometer (Difference spectroscopy)	Phosphate buffer (pH-6.8), Borate buffer (pH-9.0)	23
			UV-Visible Spectrophotometer	Methanol	24
			RP-HPLC	pH 7.0 Buffer: Methanol	25
			UV-Visible Spectrophotometer	Methanol	26
			UV-Spectrophotometer (1 st Derivative spectroscopy, Difference spectroscopy)	Phosphate buffer (pH-8.0), boric buffer (pH-9.0)	27
2.	Ethylenimine	Thiotepa	RP-HPLC	Disodium hydrogen phosphate buffer-acetonitrile (85:15V/V)	28
3.	Alkylsulphonate	Busulfan	RP-HPLC	Acetonitrile-water-tetrahydrofuran (66:32:2 V/V/V)	29
			LC-MS assay and HPLC assay	Acetonitrile	30
			RP-HPLC	Methanol-water (80:20V/V)	31
			LC-MS/MS	Acetonitrile-water (1:1 V/V)	32
			LC-MS	Ammonium acetate, formic acid, water, methanol	33
			GC	Ethyl acetate	34
			LC-MS	Acetone	35
			LC-MS	Water, methanol (80:20 V/V)	36
4.	Nitrosoureas	Carmustine	RP-HPLC	Acetonitrile-water (3:7 V/V)	37
		Lomustine	RP-HPLC	Methanol	38
			RP-HPLC	Methanol-acetonitrile	39
			LC	Potassium dihydrogen phosphate-acetonitrile	40
5.	Triazine	Dacarbazine	RP-HPLC	Methanol-phosphate buffer (2:98 V/V)	41
			RP-HPLC	Acetonitrile-0.05 M Disodium hydrogen phosphate	42
			RP-HPLC	Methanol-water (50:50 V/V)	43
		Temozolamide	RP-HPLC	Phosphate buffer-methanol (30:70 V/V)	44
			UV-Spectroscopy	Phosphate buffer	45
			RP-HPLC	Acetic acid-acetonitrile (90:10 V/V)	46

			RP-HPLC	1M HCl	47
			UHPLC-MS/MS	Ammonium formate-acetonitrile	48
			RP-HPLC	Methanol-glacial acetic acid (20:80 V/V)	49
			RP-HPLC	Ammonium acetate buffer-acetonitrile	50
			RP-HPLC	Methanol-glacial acetic acid (20:80 V/V)	51

Table II: Analytical methods for platinum coordination complexes

Sr. No.	Sub-class	Drug name	Instrument used for method development	Solvent system	Reference
1		Cisplatin	RP-HPLC	Water-methanol-acetonitrile (40:35:25 V/V/V)	52
			LC-MS/MS	Methanol	53
			UV-VIS-Spectrophotometer	Phosphate buffer (pH-7.4)	54
			RP-HPLC	Methanol-water (80:20 V/V)	55
			ICP-AES	HCl-Nitric acid (3:1 V/V)	56
			RP-HPLC	Methanol-water-acetonitrile (40:30:30 V/V/V)	57
			MECC	65 mM NaH ₂ PO ₄ - Na ₂ B ₄ O ₇ 4.0mM	58
			UV-VIS-Spectrophotometer	Phosphate buffer saline (pH-7.4)	59
			LC-Electrospray ionization tandem MS	Diethyldithiocarbamate	60
			ICP-MS	Acetonitrile-water	61
			UPLC-ESI-MS/MS	Phosphate buffer-acetonitrile	62
2		Carboplatin	LC/TOF-MS	Methanol-water	63
			UV-VIS-Spectrophotometer (Zero and 1 st order derivative)	0.1 N HCl and acetate buffer (pH-4.5)	64
			RP-HPLC	Potassium phosphate (pH-4.5)	65
3		Oxaliplatin	RP-HPLC	Purified water	66
			LC	Acetonitrile and water	67
			RP-HPLC	Methanol-acetonitrile (75:25 V/V)	68
			ICP-MS	HCl-5%, acetic acid-0.1%, thiourea-0.076%, ascorbic acid-0.01%	69
			RP-HPLC	Phosphoric acid-methanol	70

Table III: Analytical methods for antimetabolites

Sr. No.	Sub-class	Drug name	Instrument used for method development	Solvent system	Reference
1.	Folate antagonists	Methotrexate	UV-Visible-Spectrophotometer	HCl-acetonitrile	71
			RP-HPLC	pH-6.0 buffer : acetonitrile (93:7 %V/V)	72
			RP-HPLC	Water-acetonitrile-tetrahydrofuran (65:30:5 V/V/V)	73
			RP-HPLC	0.01 M Phosphate buffer-acetonitrile (89:11 %V/V)	74
			RP-HPLC	Methanol-orthophosphoric acid (70:30 V/V)	75
			UV-VIS-Spectrophotometer and HPLC	Phosphate buffer (pH-6.4)	76
			RP-HPLC	Acetonitrile-potassium dihydrogen orthophosphate (92:8 V/V)	77
			RP-HPLC	0.05 M Sodium phosphate buffer-tetrahydrofuran (95:5 V/V)	78
			RP-HPLC	Distilled water-acetonitrile with formic acid (80:20 V/V)	79
		Pemetrexed	HPLC and UV-Spectrophotometer	20mM Dibasic phosphate buffer-acetonitrile (88:12 V/V)	80
			RP-HPLC	Acetonitrile-sodium dihydrogen orthophosphate (35:65 V/V)	81
			RP-HPLC	1.0 mL Orthophosphoric acid : acetonitrile (85:15 V/V)	82
			RP-HPLC	Acetonitrile	83
			RP-HPLC	Acetonitrile-orthophosphoric acid (15:85 V/V)	84
			LC-MS	Formic acid-acetonitrile	85
2.	Purine antagonists	Azathioprine	RP-HPLC	Acetonitrile-water (50:50 V/V)	86
			RP-HPLC	Acetonitrile-water-methanol (25:70:05 V/V/V)	87
			RP-HPLC	Acetate buffer-acetonitrile-methanol (30:35:35 V/V/V)	88
			RP-HPLC	Potassium dihydrogen phosphate-acetonitrile (60:40 V/V)	89
			UV-VIS-Spectrophotometer	Methanol	90
		Fludarabine	RP-HPLC	Acetonitrile-water (10:90 V/V)	91

			RP-HPLC	Potassium dihydrogen phosphate (pH-6.0)-methanol (96:4 V/V)	92
			RP-HPLC	pH-4 Orthophosphoric acid-methanol (95:5 V/V)	93
			RP-HPLC	Perchloric acid	94
			UV-VIS-Spectrophotometer	Distilled water	95
			RP-HPLC	Methanol-water (50:50 V/V)	96
3.	Pyrimidine antagonists	Capecitabine	RP-HPLC	Phosphate buffer-acetonitrile (80:20 V/V)	97
			RP-HPLC	Methanol-ammonium acetate buffer pH-4.5 (60:40 V/V)	98
			RP-HPLC	0.01M Potassium dihydrogen phosphate-methanol (40:60 V/V)	99
			RP-HPLC	Methanol-acetonitrile-water (80:20:80 V/V/V)	100
			RP-HPLC	Methanol-buffer (70:30 V/V)	101
			RP-HPLC	Acetic acid-methanol-acetonitrile (35:60:5 V/V/V)	102
			RP-HPLC & UV-Spectrophotometer	0.1% Acetic acid-acetonitrile (35:65 V/V)	103
			UV-VIS-Spectrophotometer	Methanol	104
			UPLC & HPLC	Acetic acid-water-acetonitrile (11:2:7 V/V/V)	105
			UV-VIS-Spectrophotometer	Distilled water	106
			HPTLC	Toluene-methanol (7.5:2.5 V/V)	107
		Cytrabine	RP-HPLC	Acetonitrile-ammonium acetate(buffer) (30:70 V/V)	108
			RP-HPLC	Acetonitrile-purified water (2:98 V/V)	109
			HPLC-MS/MS	Acetonitrile-methanol	110

Table IV: Analytical methods for microtubule damaging agents

Sr. No.	Sub-class	Drug name	Instrument used for method development	Solvent system	Reference
1.	Vinca alkaloids	Vincristine	HPLC-UV-Spectrophotometer	Acetonitrile 90% in water-phosphate hydrogen phosphate buffer (32:68 V/V)	111
			UV-VIS-Spectrophotometer	Purified water	112
			RP-HPLC	Methanol	113
			RP-HPLC	0.02M Sodium dihydrogen phosphate-methanol (36:64 V/V)	114

			HPTLC	Toluene-methanol-diethylamine (8.75:0.75:0.5 V/V/V)	115
			RP-HPLC	Water-diethylamine-acetonitrile- methanol (34.9:0.1:40:25 V/V/V/V)	116
			UPLC-MS-MS	Methanol	117
		Vinblastine	RP-HPLC	Methanol-acetonitrile-ammonium acetate buffer with 0.1% triethylamine (15:45:40 V/V/V)	118
			RP-HPLC	Methanol-phosphate buffer-acetonitrile	119
		Vinorelbine	RP-HPLC	Phosphate buffer-methanol (40:60 V/V)	120
			HPLC-UV	Acetate buffer-methanol (85:15)	121
			RP-HPLC	Acetonitrile-water (60:40 V/V)	122
			RP-HPLC	Diethyl ether	123
2.	Taxanes	Paclitaxel	RP-HPLC	Acetonitrile-water (70:30, 0.1% trifluoro acetic acid)	124
			RP-HPLC	Methanol-water (80:20 V/V)	125
			RP-HPLC	Acetonitrile-water	126
			RP-HPLC	Methanol-acetonitrile-water (40:40:20 V/V/V)	127
			RP-HPLC	Phosphate buffer saline pH-7.4 with dichloromethane	128
			RP-UFLC	Acetonitrile-phosphate buffer (50:50 V/V)	129
			RP-HPLC	Acetonitrile-water (50:50 V/V)	130
			RP-HPLC	Acetonitrile-phosphate buffer (80:20 V/V)	131
			RP-HPLC	Acetonitrile-water (70:30 V/V)	132
			RP-HPLC	Acetonitrile: water (60:40 V/V)	133
			RP-HPLC	Acetonitrile-phosphate buffer (60:40 V/V)	134
3.		Docetaxel	RP-HPLC	Acetonitrile-triple distilled water (40:60 V/V)	135
			RP-HPLC	0.2% Triethylamine-acetonitrile (45:55 V/V)	136
			RP-HPLC	0.1% Ammonium acetate-acetonitrile (55:45 V/V)	137
			RP-HPLC	Acetonitrile-water (60:40 V/V)	138
			RP-HPLC	Water and acetonitrile	139
			RP-HPLC	Phosphate buffer (pH-3.6)- acetonitrile (27:73 V/V)	140
			UPLC	Acetonitrile-water (60:40V/V)	141
			ESI-MS/MS	0.1% Formic acid in methanol	142
			UV Spectrophotometer	Acetonitrile	143

Table V: Analytical methods for topoisomerase-2 inhibitors

S. No.	Sub-class	Drug Name	Instrument used for method development	Solvent system	Reference
1.		Etoposide	RP-HPLC	Acetonitrile-water (45:55 V/V)	144
			RP-HPLC	Methanol-phosphate buffer (pH-6) (80:20 V/V)	145
			RP-HPLC	Acetonitrile-acetic acid (70:30 V/V)	146
			RP-HPLC	Methanol-water	147
			RP-HPLC	Water-methanol-acetonitrile (40:35:25 V/V/V, pH 3.5)	148
			LC	Acetonitrile-water (35:65 V/V)	149
			HPLC-UV	Water-acetonitrile (70:30 V/V)	150
			RP-HPLC	Sodium acetate buffer pH-4.0-acetonitrile (70:30 V/V)	151
			HPTLC	Dichloromethane-methanol-formic acid (9.5:0.5:0.5 V/V/V)	152
		Topotecan	RP-UPLC	0.1 V/V Orthophosphoric acid in water-acetonitrile	153
			HPLC-MS	0.1% Acetic acid in acetonitrile and 0.5% acetic acid in water	154
			LC-MS/MS	0.1% Formic acid and methanol	155
			RP-HPLC & LC-MS	Methanol-isopropyl alcohol (750:250 V/V)	156
		Irinotecan	RP-HPLC	Acetic acid-acetonitrile-methanol (60:20:20 V/V/V)	157
			LC	Acetonitrile-sodium dihydrogen phosphate dehydrate buffer	158
			RP-HPLC	Acetonitrile- phosphate buffer (60:40 V/V)	159
			RP-HPLC	Phosphate buffer –acetonitrile (75:25 V/V)	160
			HPLC-MS	0.1% Acetic acid/bidistilled water and 0.1% acetic acid/ Acetonitrile	161
			RP-HPLC	Methanol-water (60:40 V/V)	162
			RP-HPLC	Acetonitrile-phosphate buffer (80:20 V/V)	163
			RP-HPLC	Water-acetonitrile (57:43 V/V)	164
			UV-VIS-Spectrophotometric	Toluene-ethyl acetate-methanol-carbon tetrachloride (9.2:5:0.9:0.8 V/V/V/V)	165

Table VI: Analytical methods for antibiotics

Sr. No.	Sub-class	Drug Name	Instrument used for Method development	Solvent system	Reference
1		Doxorubicine	RP-HPLC	Acetonitrile-ammonium hydrogen phosphate (45:55 V/V)	166
			RP-HPLC	0.05mM Ammonium acetate-methanol-acetonitrile (50:25:25 V/V/V)	167

			RP-HPLC	Acetonitrile-0.01M o-phosphoric acid (40:60 V/V)	168
			RP-HPLC	Acetonitrile-buffer (pH-3.0) (40:60 V/V)	169
			RP-HPLC	0.1%OPA-Acetonitrile (45:55 V/V)	168
			RP-HPLC	Acetonitrile-water (30:70 V/V)	169
			HPLC-MS/MS	0.05 M Ammonium acetate-acetonitrile (40:60 V/V)	170
2		Daunorubicin	RP-HPLC	Acetonitrile-methanol-sodium lauryl sulphate (9:1:10 V/V/V)	171
			RP-HPLC	Methanol-acetonitrile (75:25 V/V)	172
			RP-HPLC	0.1% OPA-acetonitrile (55:45 V/V)	173
			RP-HPLC	Methanol-phosphate buffer (60:40 V/V)	174
3		Epirubicin	RP-LC	Acetonitrile-water (30:70 V/V)	175
			RP-HPLC	Acetonitrile-methanol (80:20 V/V)	176
			RP-HPLC	Phosphate buffer-acetonitrile (50:50 V/V)	177
4		Mitomycin C	HPLC-MS/MS	Acetonitrile-isopropanol-water (20:45:35 V/V/V)	178

Table VII: Analytical methods for miscellaneous anti-cancer drugs

Sr. No.	Sub-Class	Drug Name	Instrument used for method development	Solvent system	Reference
1		Hydroxyurea	LC	Sodium acetate-acetonitrile	179
			LC	Acetonitrile-water (6.7:7.7 V/V)	180
			HPLC-UV	Ammonium acetate-acetonitrile	181
			RP-HPLC	0.2 M Sodium perchlorate-methanol (95:5 V/V)	182
2		L-asparaginase	UPLC-MS/MS	Ammonium formate- formic acid in Acetonitrile	183
3		Tretinoin	RP-HPLC	Acetonitrile-methanol (90:10 V/V)	184
			RP-HPLC	Water-glacial acetic acid (90:2 V/V)	185
			RP-HPLC	Trifluoroacetic acid-acetonitrile (15:85 V/V)	186

Microtubule damaging agents: The microtubule damaging agents are an effective category of cancer drugs with equal therapeutic benefit both in hematopoietic and solid tumours. A large number of natural agents and their analogues connect to soluble tubulin and directly to tubulin in the microtubules¹¹¹. Most of these compounds are antimetabolic agents which are able to inhibit cell growth increase by acting on the polymerization dynamics of spindles, which are necessary to proper spindle purpose of microtubules. The particular effects of MDAs on the polymer accumulation and the active constancy of the microtubules are complex¹¹⁴. Analytical methods for determination of microtubule damaging agents are discussed in Table IV.

Topoisomerase-2 inhibitors: Topoisomerase-2 inhibitors are chemical compounds used to block

the act of topoisomerase (topoisomerase-2), which is a type of enzyme that controls the changes in DNA formation by catalyzing the splitting and rejoining of the phosphodiester spine of DNA strands throughout the regular cell cycle. Topoisomerases contain sites which are well-defined targets for cancer chemotherapy treatment. It is considered that topoisomerase inhibitors block the ligation action of the cell cycle, generating specific and dual-trapped breaks that damage the reliability of the genome. Introduction of these breaks then leads to apoptosis and cell death. Analytical methods for determination of topoisomerase-2 inhibitors are summarised in Table V.

Topoisomerase-1 inhibitors: Topoisomerases are DNA enzymes that manage the topology of the supercoiled DNA double loop throughout the transcription

of reproduction of cellular genetic materials¹⁴⁴. Topoisomerase 1 inhibitors are a new group of anticancer agents with a mechanism of action intended to interrupting DNA imitation in cancer cells, the effect of which is cell death. Most, if not all Topoisomerase 1 inhibitors, are derivatives of the plant extract camptothecin. Irinotecan (CPT-11), a semi-synthetic derivative of camptothecin, is accepted in the United States for the treatment of colorectal cancer¹⁴⁶.

Antibiotics: Anticancer or antitumor antibiotics act in a manner comparable to quinolones. The major distinction among antibiotics and antineoplastic antibiotics is that the former act on bacterial cells, while the latter act on cancerous cells in the human body¹⁵⁶. Antineoplastic antibiotics affect DNA combination and reproduction by inserting into DNA strands or by producing superoxide that cause breaking of DNA strands and prevent the tumorous or cancerous cells to break up further¹⁶⁶. Analytical methods for determination of antibiotics are given in Table VI.

Miscellaneous: Miscellaneous agents are different from the major classes of cytotoxic agents. General miscellaneous agents are asparaginase and hydroxyurea. These anticancer drugs act on increased cell divisions i.e. are antiproliferative. Due to their dissimilar mechanisms, development and growth of neoplastic cells is inhibited¹⁷⁶. They also affect rapidly separating normal cells, therefore are able to repress the bone marrow, repress enlargement, damage healing, cause infertility and hair loss^{178, 181}. Analytical methods are summarized in Table VII.

CONCLUSION

Worldwide, cancer is a dangerous disease which is very significantly affecting the human population. Research and development for anticancer drugs have been the largest activity in the pharmaceutical industry in terms of the number of projects, clinical trials and spending. The different types of anticancer drugs are analyzed by using a variety of analytical methods like HPLC, GC, GC-MS, NMR, UPLC, LC-MS, Mass-Spectrophotometry, UPLC-MS/MS, LC, ESI-MS/MS, UV-VIS-Spectrophotometer and LC-UV. Analytical methodology provides analysts, researchers, industries and academicians the essential data for a specified analytical problem, sensitivity, accuracy, range of analysis and form the minimum requirement which basically is the specification of the technique. The methods which are mentioned in this review and additional methods that can be developed on this basis can be applied to diverse areas like drug testing and routine analysis in bio-analysis and quality control in pharmaceutical industries.

REFERENCES

1. Nicoliar G., Guillaume D. and Bonnabry P.: Anti neoplastic drug and their Analysis: a state of the act review. **Analyst**, 2017, 142(13), 2273-2321.
2. Shahin M., Begum K. Ayesha and Saleem M.: Analytical method for determination of anticancer drug from marine source. **IJRPC**, 2014, 4(4), 829-836.
3. Mohammad A. S., Ahmad F. and Jayanthi B.: Estimation of anticancer and antiviral drug in bulk and varied dosage forms by advanced analytical techniques: A detailed review. **WJPPS**, 2018, 7(10), 1651-1667.
4. S.Rewaria and Swamy B.M.V.: Analytical method development and validation for assay method. **IJPRS**, 2013, 2(2), 5-9.
5. Nussbaumer S., Bonnabry P. and Veuthey J. L.: LUC, Analysis of anticancer drugs: A review. Elsevier, **Talanta**, 2011, 85(5), 2265-2289.
6. Shahin M., Begum K. A. and Saleem M.: Analytical method for determination of analytical drugs from marine source. **IJRPC**, 2014, 4(4), 829-836.
7. Espinosa E. and Zamora P.: Classification of anticancer drug a new system based on therapeutic target. **Cancer Treat. Rev.**, 2003, 29, 515-523.
8. Vitthal D. D. and Milind B. V.: Development and validation of a reverse phase-HPLC method for the estimation of cyclophosphamide in bulk drug. **IJPP**, 2013, 5(2), 184-187.
9. Anilanmert B., Sertler S., Cavus F. and Cengiz S.: Validation method for occupational cyclophosphamide monitoring using LC-MS/MS and a Poroshell 120 column. **Chem. Pap.**, 2015, 69(2), 1401.
10. Zernych R. and Halkiewicz J. J.: Development and validation of SPE-HPLC-MS/MS method for determining cyclophosphamide surface water. **PJES.**, 2014, 23(5), 1537-1545.
11. Ahmad M., Usman M. and Modni A.: Development and validation of SPE-HPLC-MS/MS method for determining cyclophosphamide surface water. **Afr. J. Pharm. Pharmacol.**, 2017, 5(7), 915-922.
12. Ankalla K. K. and Gollapalli N. R.: A stability indicating UPLC method for quantification of cyclophosphamide related compound in pharmaceutical dosage form. **WJPPS.**, 2018, 7(6), 973-987.
13. Islam S., Murugan V. and Kumara P.: Development and validation of a liquid chromatographic method for the quantification of related compound in cyclophosphamide. **Pharm. Anal. Acta.**, 2018,9(9), 2-4.
14. Rastkari N., Reza A., Alehashem M. and Baniasadi S.: Rapid method for the determination of cyclophosphamide and isosphamide in urine at trace levels by Gas chromatography-mass spectrometry. **IJPCR.**, 2016, 8(5), 289-292.
15. Thangabalan B., Harini A.L. and Manoharbabu S.: Spectrophotometric estimation of cyclophosphamide in capsule dosage form. **IJPRA.**, 2015, 5(1), 36-37.
16. Torres L., Chavez-Pacheco J. L. and Naavas C.F.: A new method to quantify ifosphamide blood levels using dried

- blood spot and UPLC-MS/MS in pediatric patients with embryonic solid tumours. **PLoSOne**, 2015, 10(11), 143-421.
17. Salman D., Barton S. J. and Swinden J.: Development and validation of RP-HPLC method for the analysis of ifosfamide and mesna. **Research Gate**, 2012, 143-149.
 18. Martins I, Souza J. O. and Sanson A. L.: Simultaneous determination of cyclophosphamide and ifosfamide in plasma using SPE-HPLC-UV Method. **Lat. Am. J. Pharm.**, 2009, 28(1), 6-41.
 19. Dey S. S., Sumal B. H. and Monica P.: Method development and validation for the estimation of chlorambucil in bulk and pharmaceutical dosage forms using UV-VIS-spectrophotometric method. **J. Pharm. Res.**, 2011, 4(9), 3244-3246.
 20. Yuan Y., Painj J., Zeyu W. V. and Hui A.: Validated LC-MS/MS Method for the simultaneous determination of chlorambucil and its prodrug in mouse plasma and brain & application to pharmacokinetics. **J. Chromatogr. Sci.**, 2013, 51, 266-272.
 21. Goud E., Sasi K. and Krishna R. V.: Development and validation of RP-HPLC method for determination of related substance of bendamustine hydrochloride in bulk drug. **Der Pharmacia Sinica**, 2013, 4(1), 29-36.
 22. Bhawani S., Nageshwari H.G, Mamatha G. and Venu M.: Analytical method development and validation of bendamustine in bulk using RP-HPLC. **OAJPR**, 2018, 2(2), 000158.
 23. Annapurna M.M., Pavani S. and Anusha S.: New analytical method for the determination of bendamustine hydrochloride: An antineoplastic drug. **J. Chem. Pharm. Res.**, 2012, 4(3), 1696-1701.
 24. Padmini T. and Satyanarayan L.: Visible spectrophotometric determination of bendamustine hydrochloride in formulation sample. **World J. Pharm. Pharm. Sci.**, 2017, 6(8), 1076-1083.
 25. Pavani P., Rajeshwari T. R. and Reddy G. R.: Development and validation of stability indicating HPLC method for estimation of related substance in bendamustine hydrochloride. **Sch. Res. J.**, 2016, 8(12), 183-192.
 26. Satyanarayan L. and Padmini T.: Visible spectrophotometric determination of bendamustine by complex formation. **JST.**, 2017, 2(7), 1-6.
 27. Annapurna M. M. and Pavani S.: Derivative spectrophotometric method for the determination of bendamustine hydrochloride. **J. Appl. Pharm. Sci.**, 2012, 2(11), 139-142.
 28. Srinivas R. G., Reddy L. and Shiva K.: Novel stability indicating reverse phase HPLC method for estimation of thiotepa in its formulation. **Am. J. Pharm. Tech. Res.**, 2016, 6(4), 2249-3387.
 29. Hongxia L., G. Susan and Stair Roger K.: Comparison of LC-MS assay and HPLC assay of busulfan in clinical pharmacokinetics studies. **ISRN**, 2005, 5, 86-92.
 30. Haggie J. R, Wu Mark and Burns R. B.: Validation of a HPLC assay method for pharmacokinetic evaluation of Busulfan. **J. Chromatogr. B Biomed. Appl.**, 1997, 692(2), 437-444.
 31. Punt A. M., Langenharst J. B. and Egas A C.: Simultaneous quantification of busulfan, clorfarabine and F-ARA-A using isotope labelled standard and standard addition in plasma by LC-MS/MS for exposure monitoring in hematopoietic cell transplantation conditioning. **J. Chromatogr. B Analyt. Technol. Biomed. Life Sci.**, 2017, 6, 81-85.
 32. French D., Suijishi K. K. and Long Loyle J. R.: Development and validation of a liquid chromatography-tandem mass spectrometry assay to quantify plasma busulfan. **Ther. Drug Monit.**, 2014, 36(2), 169-174.
 33. Laenne E., Burns Robbin B. and Heggie J. R.: Gas chromatographic analysis of Busulfan for therapeutic drug monitoring. **Cancer Chemother. Pharmacol.**, 1993, 32(2), 137-142.
 34. Salamum D., Pavai M. E. and Biju G.: A rapid and sensitive LC-tandem MS method for the quantitation of busulfan level in plasma and application. **IJMR**, 2013, 137, 777-784.
 35. Yaswanth A., Murthy T.E.G.K. and Sivanath M.: Validation of RP-HPLC Analytical method for estimation of carmustine in bulk and lyophilized vials. **Indo Amer. J. Pharm.**, 2017, 2231, 55-61.
 36. Luning Z., Ling G. and Yong Z.: HPLC method validation for the quantification of lomustine encapsulated lomustine containing iohexol for CT imaging in C₆ Glioma rats. **Eur. J. Drug Metab. Pharmacokin.**, 2011, 36(2), 6-9.
 37. Shuoye Y. and Yan G.: Preparation of lomustineiohexol compound liposomes and the determination of entrapment efficiency. **J. Chem. Pharm. Res.**, 2014, 6(1), 402-407.
 38. Seshaiha Rama K. and Samanta K.: Stability indicating liquid chromatographic method for lomustine. **AGRIS**, 2015, 54(1), 213-216.
 39. Ahsanul H. and Stewart J. T.: Isocratic determination of dacarbazine and related impurities 2-azahypoxanthine and 5-amino-imidazole-4-carboxamide by HPLC on an avidin protein column. **J. Liq. Chromatogr. Relat. Technol.**, 2006, 22(16), 933-943.
 40. Zubair M. M. and Mahamood A.: Rapid and simultaneous determination of Adriamycin, bleomycin, vinblastin and dacarbazine in plasma of Hodgkin's lymphoma patients by a reversed phase HPLC Method. **J. Chil. Chem. Soc.**, 2013, 58(2), 9707-9717.
 41. Prasanath M.L., Lal and Sridhar S.: Stability indicating HPLC method for the determination of dacarbazine in pharmaceutical dosage form. **IJOP.**, 2014, 4(3), 5-12.
 42. Ishaq B. Mohammad and Prakash K. V.: Development and validation of a Reverse-phase HPLC method for analysis of temozolomide in a capsule formulation. **Int. J. Chem. Sci.**, 2013, 11(2), 1055-1063.
 43. Mohammad I. B., A. A. Hindustan and Sheikh M.: Analytical method development and validation for the estimation of temozolamide in phosphate buffer pH- 2.0 as a solvent by UV-Spectroscopy. **IRJP.**, 2014, 5(1), 6-10.
 44. Patel B. K. and Parikh R.H.: Development and validation of a RP-HPLC method for the determination of tererolomide in rats: Application to plasma

- pharmacokinetic and brain distribution study. **TOCPR.**, 2016, 8(3), 258-263.
45. Gilant E. and Kaza M.: Validation HPLC method for determination of temozolomide in human plasma. **Acta Pol. Pharm.**, 2012, 69(6), 1347-1355.
 46. MA El Mubarak and Stylos E. K., Development and validation of simple steps protein precipitation of UHPLC-MS/MS method for quantitation of temozolomide in cancer patient's plasma sample. **IPBA.**, 2019, 162,164-170.
 47. Swathimutyam P. and Bala P.: Development and validation of stability indicating RP-HPLC method for estimation of temozolomide in bulk. **WJPPS.** 2015, 4(3), 1100-1109.
 48. Daphal V. N., Holkar G. and Yadav R.: Development and validation of simultaneous determination of anastrozole and temozolomide in pharmaceutical dosage forms. **IJTAS.**, 2012, 4(2), 48-55.
 49. Palleria S. and Prabhakar B.: Development and validation of temozolomide solid lipid nanoparticle formulation by RP-HPLC method. **JPBS.**, 2015, 5(9), 745-749.
 50. Sharaf Muhammad A., Muhammad M., Faiz N. and Irshad A.: Development and validation of RP-HPLC method for the simultaneous determination of etoposide and cisplatin and its application in quality control of injectable dosage forms. **J. Chem. Soc. Pak.**, 2012, 34(2), 321-325.
 51. Shaik A. N., Altomare D. A., Leska L. J. and Trame M. N.: Development and validation of a LC-MS/MS assay for quantification of cisplatin in rat plasma and urine. **J. Chromatogr. B Analyt. Technol. Biomed. Life Sci.**, 2017, 10(46), 321-325.
 52. Siddappak K. and Hanam S.P.: Development and validation of spectrophotometric method for the determination of cyclophosphamide in bulk drug and its pharmaceutical dosage form. **IJPPS**, 2013, 5(4), 597-600.
 53. Songul T. and Filiz O.: High performance liquid chromatography determination of free cisplatin in difference cancer types. **Sch. Lib. Res.**, 2013, 5(5), 169-174.
 54. Wagle S. A., Jain D., Rathod S. and Bajaj A.: Development and validation of Inductive coupled plasma atomic emission spectroscopy analytical method for estimation of cisplatin in biological sample. **IJPER**, 2017, 51(45),783-5789.
 55. Kaushik K. Harish, Sirpuram Vijay K. and Bedada S.: A simple and sensitive validated HPLC method for quantitative determination of cisplatin in human plasma. **Clin. Res. Reg. Aff.**, 2010, 27(1), 1-6.
 56. El-Attug Mohamad, Arman A. and Elhamili A.: Development and validation of a micellarelectrokinetic capillary chromatography method for determination of cisplatin in tumor tissue. **J. Chem. Pharm. Res.**, 2013, 2(5), 308-313.
 57. Shrikhande S.S. and Jain D.S.: Development and validation of a UV-spectrophotometer method for estimation of cisplatin. **Indian Drugs**, 2014, 51(7), 23-30.
 58. Eaiming T., Li Chao and Caixing T.: Quantitative determination of platinum derivative from cisplatin in human plasma ultrafiltrate using derivatization with diethyl dithiocarbamate and LC coupled with electrospray ionization tandem Mass spectroscopy. **Anal. Methods**, 2013, 5, 7117-7126.
 59. Zhang Ti, Cai S. and Forrest W.C.: Development and validation of an inductively coupled plasma-mass spectrometry (IPC-MS) method for quantitative analysis of platinum in plasma urine and tissue. **J. Appl. Spectrosc.**, 2016, 7(9), 1529-1536.
 60. Baskarville-Abraham Irene, Boysen Gunnar and Esra M.: UPLC-ESI-MS/MS Quantitation of cisplatin 1, 2 guanine intrastand cross link. **Chem. Res. Toxicol.**, 2016, 12-15.
 61. I. Mo., PK Eggers, C. L. Raston and Lim L.Y.: Development and validation of LC-MS method for the determination of carbaplatin and paclitaxel in nanovesicles. **Anal. Bioanal. Chem.**, 2014, 406(11), 2659-2667.
 62. Rajagopal P. and Sundrarajan R.: Method development and validation of carboplatin by UV-Spectrophotometer method in bulk and pharmaceutical dosage form. **IJRAR**, 2019, 6(2), 2349-5139.
 63. Nicolas V., Sherry C. and Jason Y.: Determination of carboplatin in cannin plasma by HPLC, biomedical chromatography. **Biomed. Chromatogr.**, 2010, 24(8), 908-913.
 64. Matos Breno N., Oliverira Paula M. de and Reis Thaiene A.: Development and validation of a simple and selective analytical HPLC method for the quantification of oxaliplatin. **J. Chem.**, 2015, 70, 900-910.
 65. Ficarra R., Calabro M. L. and Cutroneo P.: Validation of a LC method for the analysis of oxaplatin in a pharmaceutical formulation using an experimental design. **J. Pharm. Biomed. Anal.**, 2002, 29(6), 1097-1099.
 66. Edla Subhashini and Sayama S. B.: RP-HPLC method for the quantification of oxaplatin. **Int. J. Sci. Invent.**, 2012, 1(1), 32-41.
 67. Morriso J. G., White P., McDougall S., Firth J. W., Woolfrey S. G., Graham M. A., and Greenslade D.: Validation of a highly sensitive ICP-MS method for the determination of platinum in biofluid: Application to clinical pharmacokinetics studies with oxaplatin. **J. Pharm. Biomed. Anal.**, 2000, 24(1), 1-10.
 68. Tasneem F., Koneru A. and Murali B.: Analytical method development and validation for residual oxaplatin are testing industrial manufacturing equipment. **EJBPS.**, 2018, 5(3), 816-869.
 69. Santos C.M. and da Costa M. Mera V.: Development and validation of spectrophotometric method for determination of methotrexate incorporated into PLGA implants. **Int. J. Drug Dev. Res.**, 2013, 5(1), 154-160.
 70. CC de Abreu, Rosa P.C., C Alives B. and La Azzalis.: Development and validation of HPLC method to determination of methotrexate in children oncology patients. **ERMPS**, 2015, 19(8), 1373-1380.
 71. Sartori T., F. Murakami S. and Pinhera C.: Development and validation of a fast RP-HPLC method for determination of methotrexate entrapment efficiency in polymeric nanocapsule. **ICS**, 2008, 46(6), 505-509.
 72. Lariya N. K. and Agrawal G.P.: Development and validation of RP-HPLC method for simultaneous determination of

- methotrexate, dexamethasone and indomethasone. **IJPPS**, 2015, 7(3), 16-22.
73. Shanmugeuel S. and Venkatachalan K.: Analytical method development and validation of layer by layer magnetic nanoparticles of methotrexate and melphalan. **WJPPS**, 2014, 3(3), 1221-1251.
 74. Suguna P. Satyanarayan B. and Chandrashekhar G.: Validation of HPLC method for the analysis of methotrexate in bulk drug and pharmaceutical dosage form. **JOCPR**, 2015, 7(9), 27-37.
 75. Ertugrul S. and Sertuglu E.: Development and validation of HPLC method for quantification of methotrexate in plasma. **AOCSI**, 2018, 4(1), 1035-1037.
 76. Roy M, Mohit M and Shah S.: Development and validation of RP-HPLC method for the determination of methotrexate in bulk and pharmaceutical dosage form. **EJPMR**, 2016, 3(5), 355-358.
 77. Patel Ankit D. and Parikh S.: Development and validation of HPLC and UV-Spectrophotometer method for estimation of pemetrexate disodium in bulk drug and pharmaceutical formulation. **IJDDR**, 2011, 3(2), 301-307.
 78. Agrawal S. K. and Rathore D.S.: Development and validation of pemetrexate by RP-HPLC method in bulk drug and pharmaceutical dosage form. **IJORPR**, 2013, 1(4), 537-542.
 79. Reddy L. Maheswara, Reddy P. R and Reddy L. B.: RP-HPLC method for the estimation of pemetrexate assay in formulation. **IJSID**, 2011, 1(1), 1-10.
 80. Banu T., Khan M.M.M. and Teja B. Bhanu.: Validated RP-HPLC Method for the determination of pemetrexate disodium in pharmaceutical dosage form. **Orient. J. Chem.**, 2010, 26(4), 1325-1332.
 81. Gupta K. R. and Balakumar C.: Development and validation of RP-HPLC method for related substance of pemetrexate disodium. **IJOPSR**, 2012, 5(2), 133-137.
 82. C. Bobbin-Dubigeon, Amaïnd M.B. and Harrenknecht C.: Development and validation of an improved LC-MS method for the determination of pemetrexate in human plasma. **JCBATBLC**, 2009, 877(24), 2451-2456.
 83. Ravishankar P., Rani A. K. and Vineela C.: Development and validation of rapid RP-HPLC method for the determination of azathioprine in bulk and pharmaceutical dosage form. **Sch. Res. Lib.**, 2015, 7(3), 85-95.
 84. Bhaskar M., Moud Md. Gayasuddin and Kulkarni V.: RP-HPLC method development for the determination of azathioprine in bulk drug and pharmaceutical dosage form. **IJCRGG**, 2010, 2(2), 1176-1179.
 85. Bandi Ramachandra P. S. and Ramanjulu C.: Validation stability indicating RP-HPLC assay method for azathioprine in pharmaceutical dosage form according to ICH Guideline. **IJPPS**, 2014, 6(10), 301-307.
 86. Jain P.S. and Surana S. J.: Development and validation of UV-Spectrophotometric method for determination of azathioprine in bulk and in formulation. **IJUPBS**, 2012, 1(2), 49-58.
 87. Mattos Ana C. de and Khali N. M.: Development and validation of an HPLC method for the determination of fluorouracil in polymeric nanoparticles. **Braz. J. Pharm. Sci.**, 2013, 41(1), 119-128.
 88. Muhammad H., Faheem A. M. and Shabana N.: Development and validation of a new HPLC method for the detection of 5-fluorouracil in mobile phase and in plasma. **Curr. Pharm. Anal.**, 2018, 14(1), 3-7.
 89. Pushpa Latha E., Reddy L. S. and Dastagiri R. Y.: Analytical method development and validation for the estimation of 5-Fluorouracil by RP-HPLC. **CJPR**, 2015, 1(4), 139-150.
 90. Usman M. M. and Mahmood A.: Development and optimization of fast and new Reverse phase HPLC method for analysis of 5-fluorouracil in human and rabbit plasma. **Pak. Vet. J.**, 2014, 35(1), 71-75.
 91. Rokad P.S. and Patil P. M.: Development and validation of analytical method for estimation of 5-fluorouracil in bulk and marketed formulation by UV-Spectrophotometer. **IJPSR**, 2017, 42(2), 8-11.
 92. Haq Nuzrul, Faiyaz S. and Fars A.: Development and validation of an isocratic sensitive and facile RP-HPLC method for rapid analysis of 5-fluorouracil and stability studies under various stress condition. **Asian J. Chem.**, 2013, 25(13), 7177-7182.
 93. Sreevastav A. S. K. and Harishababu A. K.: RP-HPLC method development and validation of capecitabine extended release tablet dosage forms. **IJPSR**, 2013, 4(11), 4477-4487.
 94. Chainitanya G., Ramana G. V. and Pawan K.M.: RP-HPLC method development and validation of capecitabine in bulk drug and formulation. **IJPAP**, 2016, 5(1), 190-198.
 95. Yashodha A., Parvathi J. and Venkataih G.: RP-HPLC method development and validation of capecitabine in pharmaceutical dosage form. **IJPAP**, 2017, 6(1), 074-092.
 96. Chettupalli A.K., Kunduru V. and Boggula N.: Development and validation of capecitabine tablet by using RP-HPLC method. **IAJPS**, 2017, 4(3), 550-557.
 97. Fatima A., Tangadpally R. and Kondepudi R. K.: Analytical method development and validation of capecitabine in bulk by RP-HPLC method. **IRIP**, 2013, 3(3), 177-180.
 98. Bhatiya M.S., Raut J. N., Barve A.C. and Patil P.S.: HPLC assay method development and validation for quantification of capecitabine in tablet and forced degradation sample. **Marmara Pharm. J.**, 2017, 21(3), 660-668.
 99. Mondal S., Reddy N. and Gosh D.: Development and validation of RP-HPLC and UV-Spectrophotometric method for the quantification of capecitabine. **IJPPS**, 2016, 8(5), 279-287.
 100. Ramesh G. and Rao M. S.: Development and validation of A simple UV-Spectrophotometric method for capecitabine assay in active pharmaceutical ingredient and its dosage form. **IJPPR**, 2015, 2(2), 152-160.
 101. Hannumanturayadu K. and Sreeramulu J.: Determination of stability indicating assay method for capecitabine in pharmaceutical drug substances a comparative study by UPLC and HPLC. **JPR**, 2012, 5(12), 5515-5519.
 102. Kumbhab S.C.R. and Salunkhe V. R.: UV-Spectrophotometric method development for capecitabine in Eudragit

- and chitosan based microspheres and its validation. **IJPBR**, 2013, 1(3), 32-38.
103. Thorat S., Chikhale R. and Rode V.: A validated stability indicating HPTLC method for the estimation of capecitabine in its tablet dosage form. **Curr. Pharm. Anal.**, 2019, 15(1), 61-66.
104. Murthy V.S.N., Rohini A. and Pravallika K.E.: Development and validation of RP-HPLC method for estimation of cytrabine in bulk and pharmaceutical dosage form. **IJPSR**, 2013, 4(12), 4573-4576.
105. Bhatnagar A., Loura S and Chaudhary M.: A stability indicating RP-HPLC method for determination of anticancer agent cytrabine in lyophilised dosage form. **EJAC**, 2012, 7(3), 160-167.
106. Hilhorst Martijn J. and Hendriks G.: HPLC-MS/MS method for the determination of cytrabine in human plasma. **Bio-analysis**, 2011, 3(14), 1285-1291.
107. Rodrigues A. S., Copes A.R., Leao A. and Couceiro A.: Development of an analytical methodology for simultaneous determination of vincristine and doxorubicin in pharmaceutical preparation for oncology by HPLC-UV. **J. Chromatogr. Sci.**, 2003, 47, 95-101.
108. Jeewantha H. A., Ivanovich S. A. and Mihailovich K. P.: Validated spectrophotometric method for the estimation of vincristine and vinblastine. **IJPPS**, 2017, 9(4), 78-86.
109. Khali H. A., El-Yazbiamad F. and Belal T. S.: High performance liquid chromatography assay for the simultaneous determination of pasaconazole and vincristine in rat plasma. **Int. J. Anal. Chem.**, 2015, 1-6.
110. Chen J., He. H., huijuan, Li S. and Shen Qi.: An HPLC method for the pharmacokinetic study of vincristine sulphate loaded PLGA-PEG nanoparticle formulation after injection of rats. **J. Chromatogr. B**, 2011, 879, 1967-1972.
111. Abib K. Amal, Sayeed A. and Jalus A. F.: Simultaneous determination of vincristine and vinblastin in *Vinca rosea* leaves by HPTLC. **IJDDR**, 2013, 5(3), 341-348.
112. Embree L., Gelmon K. A., Tolcher A.W. and Huklon N.J.: Validation of a HPLC assay method for quantification of total vincristine sulphate in human plasma following administration of vincristine sulphate liposome injection. **JPBA**, 1997, 16(4), 675-687.
113. Yang F., Wang H. and Hu Pei.: Validation of un UPLC-MS/MS method for quantitative analysis of vincristine in human urine after intravenous administration of vincristine sulphate liposome injection. **J. Chromatogr. Sci.**, 2015, 53(6), 974-978.
114. Siddiqui M.J.A., Ismail Z and Saidan Nor H., Simultaneous determination of secondary metabolites from *Vinca rosea* plant extractive by reverse phase-HPLC. **Pharmacogn. Mag.**, 2011, 7(26), 92-96.
115. Rahim R.A. and Ahmad N. H.: Determination and quantification of the vinblastine content in purple, red, and white *Catharanthus roseus* leaves using RP-HPLC method. **Adv. Pharm. Bull.**, 2018, 8(1), 157-161.
116. Gandhi B. M., Rao A. L and Rao J. V.: Development and validation of new RP-HPLC method for the estimation of vinorelbine in pharmaceutical dosage forms. **IJRPC**, 2013, 3(3), 712-716.
117. Rao A. V.: Development and validation of a simple HPLC-UV method chemotherapeutic drug in spiked human plasma. **Asian J. Pharm. Clin. Res.**, 2019, 12(1), 369-373.
118. Konda B.: Development and validation of exemestane by reverse phase HPLC. **J. Chromatogr. Sci.**, 2011, 49(8), 634-639.
119. Puozzo C., Ung H.L. and Zorza G.: A HPLC method for vinorelbine and 4-o-deacetylvinorelbine: decade of routine analysis in human blood. **J. Pharm. Biomed. Anal.**, 2017, 44(1), 144-149.
120. Jain A., Gulbanke A., Jain A. and Shilpi S.: Development and validation of the HPLC method for simultaneous estimation of paclitaxel and topotecan. **J. Chromatogr. Sci.**, 2014, 52(7), 697-703.
121. Mohammadi A., Esmaeili F., Dinvarv R. and Atyabi F.: Development and validation of a stability indicating method for the quantification of paclitaxel in pharmaceutical dosage forms. **J. Chromatogr. Sci.**, 2009, 47(7), 559-604.
122. Xia X.J., Peng J., Zhang P.X. and Jin D.J.: Validated HPLC method for the determination of paclitaxel related substances in an Intravenous emulsion loaded with a paclitaxel-cholesterol complex. **Indian J. Pharm. Sci.**, 2013, 75(6), 672-679.
123. Pallapati S., Rao T. S. and Venkat K.: Stability indicating RP-HPLC method development and validation for the estimation of paclitaxel in pharmaceutical dosage form. **WJPPS**, 2017, 6(8), 2599-2611.
124. Srinivasa M.S. and Chandan R. S.: Validated RP-UFLC method development of paclitaxel in pure and pharmaceutical dosage form. **Pharm. Method**, 2017, 8(1), 155-159.
125. Bernabeu E., Flor S., Hocht C. and Carlos T.: Development and validation of a highly sensitive HPLC method for determination of paclitaxel in pharmaceutical dosage form and biological sample. **Curr. Pharm. Anal.**, 2014, 10(3), 185-192.
126. Satyamoorthy N. K. and Rajendran V.: An approach for validated RP-HPLC method for the analysis of paclitaxel in rat plasma. **J. Appl. Pharm. Sci.**, 2014, 4(9), 073-076.
127. Tailor J. and Raul A.: Development and validation of a RP-HPLC method for *in vitro* loading and release analysis of paclitaxel coated stent. **Trends Biomater. Artif. Organs**, 2008, 22(2), 79-86.
128. Saadat I. and Ravan F.: Development and validation of rapid RP-HPLC-DAD Analysis method for simultaneous quantification of paclitaxel and lapatinib in polymeric micelle formulation. **Sci. Pharm.**, 2016, 84(2), 333-345.
129. Ahman J. and Kohli K.: Development and validation of RP-HPLC method for analysis of novel self-emulsifying paclitaxel formulation. **Research & Reviews: J. Pharm. Anal.**, 2013, 2(3), 17-27.
130. Siddaqui S., Paliwal S. and Kohil K.: RP-HPLC method for the estimation and stress degradation study of paclitaxel as per ICH guideline. **J. Chromatogr. Sep. Tech.**, 2012, 3(4), 2-4.

131. Kim D. W., Yousef A. M. and Li dong X.: Development and validation of RP-HPLC method for simultaneous determination of docetaxel and curcumin in rat plasma: Validation and stability. **Asian J. Pharm. Sci.**, 2017, 12(1), 105-113.
132. Kharkar P. B. and Talkar S.S.: A rapid and sensitive bio-analytical RP-HPLC method for the detection of docetaxel: Development and validation. **IJPER**, 2017, 51(4), 729-734.
133. Mozumdar Z. I. and Nath L.K.: A HPLC method for estimation of docetaxel in solid lipid nanoparticles. **IJPSR**, 2015, 3(8), 1058-1065.
134. laymouri S. and Varshosaz J.: Development of a rapid and precise RP-HPLC method for analysis of docetaxel in rat plasma: Application in single dose pharmacokinetic studies of folate-targeted micelles containing docetaxel. **Adv. Biomed. Res.**, 2018, 7(76), 69-73.
135. Rao B.M. and Chankraborty A.: A stability indicating HPLC assay method for docetaxel. **JPBA**, 2006, 41(2), 676-681
136. Qureshi H.K. and Mubassira A.K.N.: RP-HPLC method development and validation of docetaxel in pharmaceutical dosage form. **IJARMP**, 2018, 3(8), 14-20.
137. Khurana R. K. and Beg S.: Analytical quality by design approach for development of a validated bioanalytical UPLC method of docetaxel trihydrate. **Curr. Pharm. Anal.**, 2015, 11(3), 180-192.
138. Rafiei Pedram and Jane A.: Application of a rapid ESI-MS/MS method for quantitative analysis of docetaxel in mouse biological matrices through direct injection to mass spectrometer. **Curr. Pharm. Anal.**, 2018, 14(2), 85-94.
139. Mahawar S.: A simple ultraviolet spectrophotometric method for the estimation of docetaxel in bulk drug and formulation. **AJPA**, 2013, 3(2), 48-52.
140. Maheshbhai D. R. and Patel R. K.: RP-HPLC method development and validation of etoposide. **J. Pharm. Res.**, 2012, 5(7), 3618-3620.
141. Dhanuraju M.D.: Estimation and validation of etoposide by RP-HPLC method in rat plasma by liquid-liquid extraction. **AJPCT**, 2013, 1(2), 201-207.
142. Salono A. G. R.: Development and validation of a HPLC method for determination of etoposide in biodegradable polymeric implants. **Quim. Nova**, 2011, 35(6), 1239-1243.
143. Hayat M. M.: HPLC determination etoposide in injectable dosage forms. **J. Chil. Chem. Soc.**, 2011, 56(4), 881-883.
144. Rajagopalan R., Burchett M. and Loffoedo D.: Low- level determination of cisplatin in cleaning validation sample A high performance liquid chromatography method. **Drug Dev. Ind. Pharm.**, 2000, 20, 429-440.
145. Ashraf M., Hayat M. M., Nasim F. H. and Ahamad I.: Development and validation of RP-HPLC method for the simultaneous determination of etoposide and cisplatin and its application in quality control of injectable dosage form. **Pak. J. Clin. Soc.**, 2012, 34(2), 321-325.
146. Raja A., Akhtar A. and Shumaila B.: Rapid and sensitive liquid chromatographic method for determination of etoposide in plasma and biological samples. **J. Liq. Chromatogr.**, 2013, 36, 2796-2813.
147. Pigatto M.C., Mossmann D.L. and Dalla C. T.: HPLC-UV method for quantifying etoposide in plasma and tumor interstitial fluid by microdialysis: application to pharmacokinetic studies. **Biomed. Chromatogr.**, 2015, 29(4), 529-536.
148. Tripathi D., Mohanty R. and Bhardeaj H.: Validation, estimation and recovery studies of etoposide in bulk and injectable dosage forms using RP-HPLC and UV-Visible spectroscopy. **WJPR**, 2015, 4(5), 1956-1961.
149. Kamala B., Singh M. and Ahmad F.J.: A validated HPTLC method for the quantification of podophyllotoxin in *Podophyllum hexandrum* and etoposide in marketed formulation. **Arab. J. Chem.**, 2017, 10, 2539-2546.
150. Saini P.K., Jain C.L. and Singh R.M.: Development and validation of a RP-UPLC method for quantification of topotecan hydrochloride in bulk and injection dosage form. **IJPS**, 2010, 72(4), 494-497.
151. Rao R. N., Tripathi N. K. and Patro V. J.: Method development and validation of topotecan hydrochloride in K-2 EDTA human plasma by using HPLC coupled with tandem mass spectrometry. **Asian J. Chem.**, 2012, 24(8), 3617-3620.
152. Elena M., Posocco B. and Elisa M.: Development and validation of a high performance liquid chromatography-tandem mass spectrometry method for the simultaneous determination of irinotecan and its main metabolites in human plasma and its application in a clinical pharmacokinetics study. **PLoS One**, 2015, 1-18.
153. Mangamma K., Venkatarao D. and Mohan V.S.: Method development and validation of gemcitabine and irinotecan by RP-HPLC in pharmaceutical formulation. **Int. J. Chem. Anal. Sci.**, 2012, 3(8), 1500-1502.
154. Balaram M. V., Rao V. J. and Ramakrishna S.: Validation reverse phase HPLC method for the determination of irinotecan in pharmaceutical dosage forms. **EJC**, 2007, 4(1), 128-136.
155. Rajasekaran A., Cherukuri R. and Joghee D.: Development and validation for the determination of related substances in irinotecan HCl formulation and its stability indicating assay by RP-HPLC Method. **AJPTR**, 2013, 3(4), 646-656.
156. Raghu N.S., Reddy Y. R. and Naresh V.: Separation, identification and quantification of degradants of topotecan and its relative impurities in topotecan injection by RP-HPLC and LC-MS. **JLCRT**, 2011, 34(7), 45-51.
157. Tariq M., Negi L.M., Talegaonkar S. and Ahmad F.J.: Liquid chromatographic method for irinotecan estimation: screening of P-gp modulators. **IJPS**, 2015, 77(1), 14-23.
158. M. Vijaya J., Bhargav E. and Keerthana B.: RP-HPLC method development and validation for the simultaneous estimation of irinotecan hydrochloride and capecitabine in active pharmaceutical ingredient (APIs). **IJRPS**, 2018, 9(1), 63-67.
159. Koduru S., Tahmeena F. and Muntaha S.T.: Method development, validation and forced degradation studies of irinotecan in bulk and pharmaceutical dosage form.

- WJPPS**, 2018, 7(8), 996-1009.
160. Sharma S. and Sharma M.C.: Development and validation of spectrophotometric method and TLC densitometric determination of irinotecan HCl in pharmaceutical dosage forms. **Arab. J. Chem.**, 2016, 9, 1368-1372.
 161. Mistiran A.F., Dzarr A.A. and Gan S. H.: HPLC method development and validation for simultaneous detection of arabinoside-C and doxorubicin. **Toxicol. Mech. Methods.**, 2010, 20(8), 472-481.
 162. Kallam S.R., Srikanth J. and Prakash V.: Development and validation of stability indicating method for the quantitative determination of doxorubicin hydrochloride and its related impurities in pharmaceutical dosage forms using RP-HPLC. **J. Chem. Pharm. Res.**, 2015, 7(7), 715-724.
 163. Ashok A. H., Powar A. T., M Bhatia N. and Harinath N. M.: Development and Validation of RP-HPLC method for the determination of doxorubicin hydrochloride from vacuum foam dried formulation. **RJPT**, 2016, 9(9), 1265-1269.
 164. Manasa E., Prakash K. V., Pratap P. R. and Susena S.: Method development and validation of doxorubicin HCl in API and its formulation by spectrophotometer. **IJPCBS**, 2013, 3(4), 1006-1009.
 165. Renikunta M., Reddy T.R. M. and Konde A.: RP-HPLC method development and validation for estimation of doxorubicin and in bulk and pharmaceutical dosage form. **IJAMSCR**, 2019, 7(1), 9-16.
 166. Dharmalingam S. R., R. Srinivasan, Chidambaram K. and Nadaraju S.: A simple HPLC bioanalytical method for the determination of doxorubicin hydrochloride in rat plasma: application to pharmacokinetic studies. **Trop. J. Pharm. Res.**, 2014, 13 (3), 409-415.
 167. Park J. S., Hye-kyung K. and Lee Hye-W.: Validation of HPLC-MS/MS Method for determination of doxorubicin in mouse serum and its small tissues. **KJCP**, 2006, 6, 20-32.
 168. Reddy Lakki R.H., Meda N. and Murthy R. R.: Rapid and sensitive HPLC method for the estimation of doxorubicin in dog blood- the silver nitrate artefact. **Acta Pharm.**, 2005, 55, 81-91.
 169. Srivastava A.K., Pallavi K., Shamshul S. and Khurana S.C.: Development of stability indicating HPLC method for the assay of epirubicin in the presence of degradants, **ACAIJ**, 2011, 10(10), 670-675.
 170. Lukawska M., Oszczapowicz I. and Jelinska A.: Development and validation of RP-HPLC Method for determination of novel derivatives of daunorubicin. **Chem. Anal.**, 2009, 54, 907.
 171. Pallapati S. and Rao T.S.: Development and validation of a stability-indicating RP-HPLC method for estimation of daunorubicin- A chemotherapy drug in bulk and pharmaceutical formulation. **WJPPR**, 2017, 6(7), 1158-1174.
 172. Sujana K. and Satyanarayana V.: Validated stability indicating RP-HPLC method for simultaneous estimation of daunorubicin and cytarabine in bulk and its pharmaceutical dosage for. **IJPSR**, 2017, 59.
 173. Yang Y.: Development and validation of a high-performance liquid chromatography – tandem mass spectrometric method for quantification of daunorubicin in rat plasma. **J. Talanta**, 2007, 71(2), 596-604.
 174. Kushagra R.: Method development and validation of RP-HPLC method for simultaneous estimation of daunorubicin and cytarabine in synthetic mixture. **WJPPS**, 2018, 7(5), 1301-1310.
 175. Kurbanoglu S. and Gumutas M.: Development and validation of a stability indicating RP-HPLC method for the determination of anticancer drug epirubicin in pharmaceuticals. **JLCRT**, 2014, 37(11), 1583-1596.
 176. Tariq M., Thomas S. and Singh A.: Developed and validated stability indicating HPLC method for the determination of epirubicin in bulk drug, marketed injection and polymeric nanoparticles. **Braz. J. Pharm. Sci.**, 2019, 54(4).
 177. Sreedevi A., Lakshman Rao A. and Kalyani L.: Stability indicating HPLC Method for analysis of epirubin in pharmaceutical dosage form. **Indo Am. J. Pharm. Res.**, 2013, 3(10), 8249-8259.
 178. Hymer C. B. and Connor T. H.: Validation of an HPLC-MS/MS and wipe procedure for mitomycin C contamination. **J. Chromatogr. Sci.**, 2015, 53(4), 619-624.
 179. Pujari M.P. and Barrientos A.: Determination of hydroxyurea in plasma and peritoneal fluid by HPLC using electrochemical detection. **JCBBSA**, 1997, 694(1), 185-191.
 180. Legrand T., Rokatoson M.G. and Galacteros F.: Determination of hydroxyurea in human plasma by RP-HPLC using derivatization with xanthidrol. **J. Chromatogr. B Analyt. Technol. Biomed. Life Sci.**, 2017, 1(1064), 85-91.
 181. Elias Darcielle B. D.: Standardization method for measurement of hydroxyurea by Ultra high efficiency liquid chromatography in plasma of patients with sickle cell disease. **Braz. J. Pharm. Sci.**, 2014, 50(3).
 182. Gordon A., Asante S.B., Bukari Y. and Gati L.: Method development and validation for hydroxyurea in fixed oral solid dosage forms using RP-HPLC with DED detection. **RAJAR**, 2015, 1(7), 235-242.
 183. Jeong H. C. and Kim T.: Development of a UPLC-MS/MS method for the therapeutic monitoring of L-asparaginase. **Transl. Clin. Pharmacol.**, 2018, 26(3), 134-140.
 184. Sheliya K., Shah K. and Kapupara P.: Development and validation of analytical method for simultaneous estimation of mometasone furoate, Hydroquinone and tretinoin in topical formulation by RP-HPLC. **J. Chem. Pharm. Res.**, 2014, 6(4), 934-940.
 185. Roy C. and Chakrabarty J.: Stability indicating RP-HPLC method development and validation for determination of potential degradation impurities of tretinoin in tretinoin topical pharmaceutical formulation. **Der Pharmacia Sinica.**, 2013, 4(4), 6-14.
 186. Sheliya K., Shah K. and Kapupara P.: Development and validation of analytical method for simultaneous estimation of mometasone furoate, hydroquinone and tretinoin in topical formulation by RP-HPLC. **J. Chem. Pharm. Res.**, 2014, 6(4), 934-940.